Climate change impacts on genetically differentiated *Telopea speciosissima* (NSW Waratah) coastal and upland populations

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Summary

Changes in key climatic variables (e.g., atmospheric CO2, air temperature and water availability) are occurring at unprecedented rates and having substantial impacts on functionality, biodiversity and productivity of terrestrial ecosystems. Because forests dominate terrestrial net primary production and play a prominent role in the global carbon cycle, understanding the capacity of woody species to cope with simultaneously changing climatic variables is critical for the management of natural resources and the conservation of biodiversity. One fundamental way that plants may respond to rapid climate change in the short-term is to adjust their growth and physiology via phenotypic plasticity – the ability of a genotype to express multiple phenotypes in response to environmental change, which is thought to be particularly important for woody species with long generation times. For any given species, plant populations originating from different environments usually differ in their responses to the same environmental change, as evidence of intraspecific variation in phenotypic plasticity. Although some progress has been made on intraspecific variation in woody plant response to climate change, no studies have looked into the interactive effects of concurrently changing climatic variables on their intraspecific variation in phenotypic plasticity. Therefore, this project was designed to assess the impacts of key climatic variables (i.e., [CO2], temperature, and water availability) on growth and physiology of woody plant populations originating from contrasting environments, with a focus on the intraspecific variation in their capacity to cope with climate change. *Telopea speciosissima* (Proteaceae; Shrub; open woodland), consisted of two populations originating from climatically differentiated regions. Treatment levels (i.e., changes in [CO2], temperature, and water availability) in this research were chosen based on predicted climatic conditions within this century. The goal was to use this woody species to generate

In the first experiment, the main and interactive effects of elevated [CO2] (CE) and elevated temperature (TE) on growth and physiology of the Coastal (warmer, less variable temperature environment) and the Upland (cooler, more variable temperature environment) genotypes of *T. speciosissima* were assessed. Seedlings were grown under two [CO2] (400 μ l l-1 and 640 μ l l-1) and two temperature (26/16 °C and 30/20 °C for day/night) treatments. Both genotypes were positively responsive to CE (35% and 29% increase in whole-plant dry mass and leaf area, respectively), but only the Coastal genotype exhibited positive growth responses to TE. It was observed that the Coastal genotype (no change in dry mass or leaf area). No intraspecific variation in physiological plasticity was detected under CE or TE, and the interactive effects of CE and TE on intraspecific variation in phenotypic plasticity were also largely absent. Overall, TE

was a more effective climate factor than CE in exposing genotypic variation in this woody species. Results from the chapter contradict the paradigm that genotypes from more variable climates will exhibit greater phenotypic plasticity in future climate regimes.

In the second experiment, the main and interactive effects of elevated [CO2] (CE) and elevated temperature (TE) on growth and physiological responses to drought of the Coastal (warmer and relatively wetter environment) and the Upland (cooler and relatively drier environment) genotypes of *T. speciosissima* were investigated. Seedlings were grown under two [CO2] (400 µl l-1 and 640 µl l-1) and two temperatures (26/16 °C and 30/20 °C for day/night). During the period of experiment, half of the seedlings were supplied with full watering (i.e., the well-watered treatment), while the other was subjected to controlled drought/recovery cycles (i.e., the drought treatment). The two genotypes showed similar declines in growth and photosynthesis under drought conditions across [CO2] and temperature treatments, and did not exhibit differences in response to drought stress. Regardless of genotype, TE negatively affected plant drought resistance by accelerating the process of drought seedlings becoming physiologically stressed, while CE did not influence the capacity of plant drought response. Overall, these results suggest that woody plant populations originating from different environments may not necessarily show intraspecific variation in response to drought under current or predicted future climates. These findings also indicate that temperature is likely to be a stronger determinant than [CO2] in affecting woody plant response to drought in the context of climate change.

In conclusion, the research addressed the main and interactive effects of changes in multiple climatic variables (i.e., [CO2], temperature, and water availability) on growth and physiology of *Telopea speciosissima*, with a focus on the intraspecific variation in their responses between populations originating from different environments. Results of this research were reported based on the treatment levels chosen for the experiments. Significant intraspecific variation in growth plasticity when responding to a constant mild warming (TE; ambient + 3.5–4.0 °C) was found. In contrast, populations did not differ in their growth or photosynthetic responses to elevated [CO2] (CE) or to sustained drought. These results together suggest that temperature would be more effective than [CO2] or water availability in exposing intraspecific variation in phenotypic plasticity for woody plant populations under future climates. The predicted relationships between phenotypic plasticity and source environment variability of plant populations was not supported by the findings from *T. speciosissima*, indicating that woody plant populations originating from more variable environments may not necessarily show greater phenotypic plasticity in response to climate change. This work expands current knowledge regarding the interactive effects of simultaneously changing climatic variables on woody plant growth and physiology.

More importantly, this research contributes valuable information on intraspecific variation in phenotypic plasticity of woody plant populations in response to changing climatic variables, as well as the association between phenotypic plasticity and source environment variability, which will assist in making robust predictions of the distribution and abundance of woody species under future climates.

Introduction

Increasing emissions of greenhouse gases from anthropogenic activities including rapid fossil fuel consumption and land use changes are contributing to the ongoing global climate change. Atmospheric carbon dioxide concentrations ($[CO_2]$) have been increasing from about 280 µl l⁻¹ before the industrial revolution to over 400 µl l⁻¹ nowadays and are projected to exceed 550–900 µl l⁻¹ by the end of this century (Collins *et al.*, 2013). Rising $[CO_2]$ is expected to cause a 0.3–4.8 °C increase in the global mean air temperature during same time period (Solomon *et al.*, 2009; Collins *et al.*, 2013). Embedded with this

climatic warming trend, increases in the frequency and intensity of extreme climatic events such as drought and heat waves are also anticipated through this century according to current climate change models (Meehl & Tebaldi, 2004; Della-Marta *et al.*, 2007; Kharin *et al.*, 2007; Ballester *et al.*, 2010; Yao *et al.*, 2013). Similar predictions have been made for Australia in terms of climate change. By 2070, annual mean air temperatures in Australia are projected to increase by 1–6 °C, with summer temperatures exceeding 35 °C expected to occur over 10 times more frequently in the meantime (Pearce *et al.*, 2007). Annual precipitation is also predicted to decline in many parts of Australia in the coming decades (Pittock, 2003; Pearce *et al.*, 2007; Moise & Hudson, 2008).

Changes in these climatic variables are likely to substantially regulate plant growth, function and development, thereby affecting functionality, biodiversity and productivity of terrestrial ecosystems (Nemani *et al.*, 2003; Ciais *et al.*, 2005; Williams *et al.*, 2008; Allen *et al.*, 2010; Matesanz *et al.*, 2010; Barnosky *et al.*, 2012). On the global scale, forests cover about 30% of land surface and dominate terrestrial net primary production (up to *c.* 70%), playing a prominent role in the global carbon cycle (Schimel *et al.*, 2001; Karnosky, 2003; Norby *et al.*, 2005; FAO, 2006; Bonan, 2008; Beer *et al.*, 2010; Pan *et al.*, 2011). Therefore, quantifying and understanding the capacity of woody species to cope with simultaneously changing climatic variables is of particular importance for the management of natural resources and the conservation of biodiversity (Sala *et al.*, 2000; Mawdsley *et al.*, 2009).



To cope with the ongoing rapid anthropogenic climate change, species will have to rely on different approaches such as distinct ecological (e.g., habitat shifts and phenotypic plasticity) and evolutionary strategies (e.g., adaptation and gene flow), as well as in combination (Kawecki, 2008; Anderson *et al.*, 2012). One fundamental way that plant species may respond to changing climatic variables in the short-term is to adjust their growth and physiology via phenotypic plasticity – the ability of a genotype to express multiple phenotypes in response to environmental change (Bradshaw, 1965; Sultan, 2000; Nicotra *et al.*, 2010; Anderson *et al.*, 2012). For woody plant species with long generation times, phenotypic plasticity is thought to be particularly important for acting as a buffer against rapid climate change and providing growth advantages (Valladares *et al.*, 2007; Chevin *et al.*, 2010; Nicotra *et al.*, 2010), because their evolutionary responses by natural selection might be too slow to mitigate the effects of rapid environmental change.

For any given plant species, when genotypes show differentiated responses to the same environmental change, intraspecific variation in phenotypic plasticity exists, known as significant genotype (G) by environment (E) interactions (Nicotra *et al.*, 2010; Aspinwall *et al.*, 2015). Intraspecific variation in phenotypic plasticity would not only influence the habitat range occupied by plant species, but also affect the ecological and evolutionary responses of plant species to changing environments (Sultan, 2000; Van Kleunen & Fischer, 2005; Valladares *et al.*, 2007; Williams *et al.*, 2008; Nicotra *et al.*, 2010; Aspinwall *et al.*, 2015). For instance, genotypes with low phenotypic plasticity may tolerate and persist under extreme conditions to survive and maintain growth (Schlichting, 1986; Thompson, 1991), while genotypes with high phenotypic plasticity may be capable of rapid resource uptake and show increased growth when conditions are optimal (Grime & Mackey, 2002). Therefore, studies on intraspecific variation in phenotypic plasticity of woody plants in response to changing climates are essential for making robust predictions of woody species responses under global climate change, as well as identifying genotypes that exhibit the capacity to increase or maintain productivity under more extreme climatic conditions in the future (Nicotra *et al.*, 2010; Aspinwall *et al.*, 2015; Moran *et al.*, 2016).

Although previous studies have demonstrated intraspecific variation in growth or physiological plasticity of woody plant species in response to elevated [CO₂] (e.g., Dickson *et al.*, 1998; Mohan *et al.*, 2004; Cseke *et al.*, 2009), or elevated temperature (e.g., Weston & Bauerle, 2007; Weston *et al.*, 2007; Drake *et al.*, 2015), or water deficit (e.g., Cregg & Zhang, 2001; Monclus *et al.*, 2006; Ramirez-Valiente *et al.*, 2010; Bansal *et al.*, 2015), the nature and basis of intraspecific variation in phenotypic plasticity within woody species under climate change remains largely unknown. To date, no study has looked into the interactive effects of concurrently changing climatic variables on intraspecific variation in phenotypic plasticity of woody plant species. To better understand and predict how woody plants would respond to future climatic scenarios, manipulations of combinatorial experiments assessing the main and interactive effects of [CO₂], temperature and water availability on intraspecific variation of woody plant responses are necessary.

The overall objective of this research was to assess the impacts of key climatic variables (i.e., $[CO_2]$, temperature, and water availability) on growth and physiology of woody plant populations originating from contrasting environments, with a focus on the intraspecific variation in their capacity to cope with climate change. The ecologically and economically important Australian native woody species, *Telopea speciosissima* (Proteaceae; Shrub; open woodland), consisting of two populations originating from climatically differentiated regions. The research was conducted in a state-of-the-art glasshouse facility located at the University of Western Sydney with pot-grown woody plant seedlings. The glasshouse was set to control $[CO_2]$ (ambient and ambient + 240 µl Γ^1) and temperature (ambient and ambient + 4.0 °C) conditions for simulating current and future climatic scenarios within this century based on model predictions. These combinatorial studies on woody species representing varying taxa and functional attributes were aimed to improve understanding on intraspecific variation of woody plant growth and physiological responses to simultaneously changing climatic variables (i.e., $[CO_2]$, temperature, and water availability). Specifically, addressing the following questions:

- (1) Do changes in climatic variables independently or interactively expose intraspecific variation in phenotypic plasticity of woody plant populations originating from different environments?
- (2) If differentiated responses between woody plant populations exist, what are the relationships between phenotypic plasticity and their source environmental variability?
- (3) How will climatic variables interactively affect growth and physiology of woody plants under future climates?

Methodology

Telopea speciosissima R.Br. (Proteaceae), commonly known as the Waratah (Weston & Crisp, 1994), is an endemic woody species (and New South Wales floral emblem) in the Sydney Bioregion of Australia. This species occurs sporadically in small populations across a range of climatic and altitudinal zones, and generally flowers over a six-week period in spring (September – October in warmer areas, but later in cooler areas), followed by a vegetative flush of growth (Nixon, 1997). A previous study on morphology and population genetics of *T. speciosissima* has revealed three distinct gene pools (coastal, upland and southern) among natural populations; the coastal and upland gene pools mix at mid-elevations along an altitudinal gradient (Rossetto *et al.*, 2011). Distinction in climate between habitats of coastal and upland gene pools is mainly characterized by differences in air temperature and precipitation. The coastal region is warmer and wetter than the upland region, but the latter experiences greater levels of temperature variability (Table 2-1). Thus, *T. speciosissima* is well suited for studying the association between phenotypic plasticity and source environment variability of genetically differentiated woody plant populations.

Results from such studies will provide useful information on the importance of intraspecific variation in phenotypic plasticity in determining woody species growth and physiology under climate change.

Plant material and growing conditions

Seeds were collected in May-June 2012 from two T. speciosissima populations along the elevational gradient, which are from coast to upland gene pools. At <u>least 10 waratah mother plants from each population were selected for the glasshouse</u> experiment. Seeds were directly sown into pots and cultivated in four adjacent, naturally lit glasshouse compartments ($3.0 \text{ m} \times 5.0 \text{ m} \times 3.5 \text{ m}$, width × length × height each) which are [CO2] and temperature controlled (more details are provided in Ghannoum et al. 2010). Of the four glasshouse compartments, two are set to simulate the ambient temperature (26/18 °C for day/night; ambient temperature treatment) and the other two are programmed to simulate a constant 4 °C increase in temperature (30/22 °C for day/night; high temperature treatment). Within each temperature treatment, plants were grown at ambient [CO2] (target 400 µL L-1) and elevated [CO2] (target 640 µL L-1).

 Table Error! No text of specified style in document.-1 The 40-year (1971–2010) summary of precipitation and air temperature in the coastal (180 m altitude)

 and upland (1150 m altitude) regions, from which the *Telopea speciosissima* Coastal and Upland genotypes were sampled for this study

Coastal	region		Upland		
Mean	Range	CV	Mean	Range	CV

Precipitation (mm)

Annual	1243	792–2044	0.266	856	393-1265	0.255
Summer	372	146–946	0.458	276	53–539	0.381
$T_{max}(^{\circ}C)$						
Annual	22.8	21.9–23.8	0.159	18.5	17.0-20.0	0.296
Summer	26.9	24.1-29.8	0.043	24.9	20.8–29.1	0.069
T_{min} (°C)						
Annual	13.2	12.2–14.0	0.319	7.4	6.2-8.4	0.585
Summer	18.1	15.9–20.4	0.054	12.6	9.4–15.3	0.095

Range refers to the minimal and maximal values of annual/summer Means. CV, coefficient of variation, defined as the ratio of the standard deviation to the mean; T_{max} , maximum air temperature; T_{min} , minimum air temperature. CVs for precipitation were calculated based on the annual/summer means (n = 40); while CVs for temperature were first calculated based on the monthly means within each year (n = 12) or summer (n = 3), and then averaged across 40 years. Air temperatures selected as the reference for T_A in the experiment are shown in bold.

Drought treatment

After seed germination and seedling growth, successfully established seedlings from each population were randomly selected for conducting a sustained drought experiment. Half of the selected seedlings were watered daily to field capacity (well watered treatment), while the other half of seedlings underwent two sustained periods of water stress (3 - 4 weeks for each period; sustained drought treatment). In addition, we re-watered the drought-treatment plants

following extensive drought to field capacity (i.e. recovery phase). Drought was achieved by initial cessation of daily watering and followed by controlled additions of small amounts of water on a daily basis to maintain low stomatal conductance.

Growth measurements

Seedling germination rates, seedling growth and survivorship were recorded for several months prior to the drought treatment. Seedling height, leaf number, area and weight, and stem and root weights for each treatment were measured throughout the project to calculate the following growth indices: specific leaf weight (SLW), leaf weight ratio (LWR), leaf area ratio (LAR) and shoot/root weight ratio (SRR). Additional leaves will be used to analyse leaf N and P concentrations, and total nonstructural carbohydrates (TNCs; sugars and starch).

Plant physiological traits

Gas exchange measurements were conducted on attached, recently fully expanded leaves using the Li-Cor 6400 portable photosynthesis system (Li-Cor Inc. USA). Measurements of net photosynthesis at saturating light (Asat), stomatal conductance (gs), the ratio of intercellular to ambient CO2 concentration (Ci/Ca) and leaf water use efficiency (WUEL) were conducted before measuring the responses of photosynthetic assimilation rates to intercellular CO2 concentrations (A/Ci curves) and the responses of assimilation rates to leaf temperature (A/TL curves). Photosynthetic responses of each treatment will be monitored on a weekly basis during the whole experimental period. A/Ci curves within different water availability phase (i.e. pre-drought, drought, and recovery phases) were also determined.

All data were analysed using a general linear model, factorial analysis of variance (ANOVA) with three main factors – genotype, growth [CO2] and growth temperature, with two levels within each factor. The effect of ontology was also tested with the addition of plant dry mass as a covariate in the analyses, but overall there was no change from the original analyses. Tukey's HSD tests were used to compare means for both genotypes among the [CO2] and temperature treatments (see Tables 2-3 and 2-4). Relationships between whole-plant dry mass and other parameters were analysed using linear regression analysis. Data were log-transformed when necessary to meet assumptions of homoscedasticity and normality. Results were considered significant in all cases if P < 0.05. All analyses were performed in R (version 3.1.0; R Foundation for Statistical Computing, Vienna, Austria).

Results

PART 1: Plant growth and physiology

Plant growth and dry mass allocation

Whole-plant dry mass and leaf area varied significantly between genotypes and were both affected by growth $[CO_2]$ and temperature (Fig. 2-1a and 1b; Table 2-2 and 2-3). Overall, the Coastal genotype was more productive and possessed higher leaf area, compared to the Upland genotype. Across genotypes, C_E increased whole-plant dry mass and leaf area by 35% and 29%, respectively. T_E also increased whole-plant dry mass and leaf area in the Coastal genotype by 47% and 85%, respectively, but did not significantly affect either trait in the Upland genotype, indicating genotypic variation in growth responses to temperature. LMA was higher under C_E and in the Upland genotype, but did not vary with temperature (Fig. 2-1c; Table 2-2 and 2-3). LAR did not show differences between genotypes or vary with [CO₂], but increased 24% under T_E (Fig. 2-1d; Table 2-2 and 2-3).

Dry mass allocation to different plant organs differed between genotypes, with allocation varying strongly with temperature (Fig. 2-2; Table 2-2 and 2-3). Across genotypes and $[CO_2]$ treatments, T_E increased leaf and stem mass fractions, but decreased tuber and root mass fractions, thereby generating a 50% reduction in the mean Root/Shoot ratio. Compared to the Upland genotype, the Coastal genotype allocated more dry mass to below-ground organs (higher fraction of tuber and root mass; Fig. 2-2c and 2d), but less dry mass to above-ground organs (a lower fraction of leaf mass; Fig. 2-2a), resulting in significantly higher ratios of Root/Shoot (Fig. 2-2e). The fraction of stem mass did not vary between genotypes; the Coastal genotype showed a 43% increase in stem mass fraction under T_E , but no change occurred in the Upland genotype, suggesting a significant genotype × temperature interaction (Fig. 2-2b and Table 2-2). C_E did not affect dry mass allocation.

Leaf gas exchange

In general, genotypes did not differ in leaf gas exchange parameters. However, C_E and T_E significantly affected all photosynthetic parameters, except g_s and A/T_L parameters (Table 2-2 and 2-3). Photosynthesis (A_{sat}) was 30% higher in $C_E T_A$ and 19% higher in $C_E T_E$ compared with the C_A treatments (Fig. 2-3a). Stomatal conductance (g_s) was not affected by C_E or T_E , although there was a significant interaction between genotype and temperature (Fig. 2-3b). Across temperature treatments, photosynthetic capacity traits (A_{max} , V_{cmax} and J_{max}) decreased by c. 20% under C_E (Fig. 2-3c, 3d and 3e). Growth temperature had little effect on A_{max} or V_{cmax} , but J_{max} was significantly reduced by c. 16% under T_E ; consequently, there was an 8% decline (on average) in J_{max}/V_{cmax} under T_E (Fig.

2-3f). $C_{\rm E}$ alone had no significant effect on $J_{\rm max}/V_{\rm cmax}$, but a 15% decrease in $J_{\rm max}/V_{\rm cmax}$ was observed under $C_{\rm E}T_{\rm E}$, suggesting a significant interaction between [CO₂] and temperature (Table 2-2 and 2-3). Photosynthetic thermal optimum ($T_{\rm opt}$) and light-saturated photosynthesis at thermal optimum ($A_{\rm opt}$) did not differ between genotypes or vary between growth temperatures, but increased under $C_{\rm E}$ by an average of 8% and 26%, respectively (Fig. 2-4; Table 2-2 and 2-3). The average increase of $T_{\rm opt}$ was 2.4 °C for the Coastal genotype and 1.9 °C for the Upland genotype, respectively. The main and interactive effects of genotype, [CO₂], and temperature had little effect on $A/T_{\rm L}$ parameters, except for a marginally significant interaction between genotype and temperature on parameter C (Table 2-2).

Table Error! No text of specified style in document.-2 Main and interactive effects of genotype, $[CO_2]$ and temperature on growth, photosynthetic and carbohydrate parameters of two *Telopea speciosissima* genotypes grown at two $[CO_2]$ and two temperatures

	Main eff	fects		Interaction	S			
							Genotype ×	
				Genotype	Genotype ×	$[CO_2] \times$	$[CO_2] \times$	
Parameter	Genoty	pe [CO ₂]	Temperatu	re $\times [CO_2]$	Temperat	ture Tempe	erature Tempera	ature
Growth	0.000	0.004	0.033	0.452	0.024	0.406		
Whole-plant DM (g)	0.000	0.004	0.035	0.452	0.024	0.400	0.511	
Leaf Area (cm ²)	0.000	0.013	0.000	052	0.008	0.720	0.338	
LMA (g m^{-2})	0.024	0.025	0.346	0. 90	0.467	0.391	0.128	

Table 2-2 (continued)

Main effects				Interactions			
							Genotype ×
				Genotype	Genotype ×	$[CO_2] \times$	$[CO_2] \times$
Parameter	Genotype	[CO ₂]	Temperature	× [CO ₂]	Temperat	ture Tempera	ature Temperature
$-LAR (m^2 kg^{-1})$	0.129	0.658	0.000	-0.330	0.364	0.353	0.622
Leaf mass fraction	0.000	0.268	0.000	0.638	0.693	0.650	0.780
Stem mass fraction	0.095	0.270	0.000	0.221	0.014	0.649	0.375
Tuber mass fraction	0.000	0.071	0.000	0.781	0.063	0.379	0.607
Root mass fraction	0.012	0.615	0.000	0.239	0.271	0.860	0.748
Root/Shoot ratio	0.000	0.507	0.000	0.408	0.401	0.580	0.985
Leaf gas exchange							
$A_{\rm sat}$ (µmol m ⁻² s ⁻¹)	0.961	0.000	0.021	0.344	0.647	0.325	0.605
$g_{\rm s} ({\rm mol}\;{\rm m}^{-2}\;{\rm s}^{-1})$	0.247	0.836	0.304	0.941	0.020	0.367	0.081
$A_{\rm max} (\mu { m mol} { m m}^{-2} { m s}^{-1})$	0.374	0.000	0.106	0.363	0.394	0.101	0.628
$V_{\rm cmax} \ (\mu { m mol} \ { m m}^{-2} \ { m s}^{-1})$	0.175	0.000	0.080	0.897	0.446	0.319	0.740
$-J_{\text{max}} (\mu \text{mol m}^{-2} \text{ s}^{-1})$	0.177	0.000		-0.440	0.301	0.884	0.688
0.000				0.583	0.808	0.029	0.099
$J_{\rm max}/V_{\rm cmax}$	0.614	0.059	0.002				
A/T_L	0.702	0.384	0.387	0.477	0.277	0.785	0.902
Parameter A				0.393	0.164	0.590	0.914
Parameter B	0.732	0.067	0.302	15			

	Main effect	ts		Interaction	S		
							Genotype ×
				Genotype	Genotype ×	$[CO_2] \times$	[CO ₂] ×
Parameter	Genotype	[CO ₂]	Temperature	× [CO ₂]	Tempera	ture Temper	ature Temperatu
Parameter C	0.486	0.104	0.141	0.214	0.043	0.584	0.778
$T_{\rm opt}(^{\circ}{\rm C})$	0.837	0.000	0.380	0.666	0.252	0.194	0.677
$A_{\rm opt}$ (µmol m ⁻² s ⁻¹)	0.578	0.000	0.932	0.929	0.774	0.318	0.455
Carbohydrates							
Whole-plant St (mg g ⁻¹)	0.031	0.020	0.000	0.891	0.507	0.542	0.385
Whole-plant Ss (mg g ⁻¹)	0.004	0.149	0.086	0.605	0.105	0.879	0.830
Whole-plant NSC (mg g ⁻¹)	0.015	0.049	0.033	0.570	0.399	0.504	0.584
Leaf St (mg g ⁻¹)	0.338	0.002	0.000	0.763	0.810	0.871	0.133
Stem St (mg g^{-1})	0.002	0.723	0.000	0.317	0.549	0.787	0.772
_Tuber St (mg g^{-1})	0.000	0.234	0.004	_0.052	0.597	0.675	0.677
Root St (mg g^{-1})	0.014	0.665	0.136	0.637	0.262	0.288	0.165
Leaf Ss (mg g^{-1})	0.000	0.702	0.540	0.406	0.164	0.115	0.293
Stem Ss (mg g ⁻¹)	0.273	0.043	0.173	0.640	0.743	0.074	0.420
Tuber Ss(mg g ⁻¹)	0.278	0.002	0.143	0.709	0.013	0.135	0.248
Root Ss (mg g^{-1})	0.019	0.000	0.000	0.373	0.290	0.014	0.005

Table 2-2 (continued)

DM, dry mass; LMA, leaf area per mass; LAR, leaf area ration; St, starch; Ss, soluble sugars; NSC, non-structural carbohydrates. P-values from the three-

way ANOVA are presented, based on ten replicates (n = 10) for growth parameters and five replicates (n = 5) for the others. Significant values (P < 0.05) are shown in bold. **Table** Error! No text of specified style in document.-3 Summary of means for growth, photosynthetic and carbohydrate parameters of *Telopea speciosissima*Coastal and Upland genotypes grown under the four $[CO_2]$ and temperature treatments, as described in the Materials and methods

		Treatment			
Parameter	Genotype	$C_{\mathrm{A}}T_{\mathrm{A}}$	$C_{\rm A}T_{\rm E}$	$C_{\rm E}T_{\rm A}$	$C_{\rm E}T_{\rm E}$
Growth					
Whole-plant DM (g)	Coastal	6.3 ± 0.9^{bcd}	9.2 ± 1.0^{ab}	8.0 ± 1.1^{abc}	$11.7\pm1.5^{\rm a}$
	Upland	4.5 ± 0.2^{cd}	4.3 ± 0.8^{d}	6.1 ± 0.8^{bcd}	6.5 ± 0.6^{abcd}
Leaf Area (cm ²)	Coastal	$267 \pm 46^{\circ}$	524 ± 64^{ab}	322 ± 57^{bc}	559 ± 64^a
	Upland	$205\pm14^{\circ}$	231 ± 44^{c}	$283\pm43^{\circ}$	350 ± 31^{abc}
LMA (g m ⁻²)	Coastal	$98.5\pm3.8^{\mathrm{a}}$	92.3 ± 2.1^a	$102.0 \pm 4.9^{\mathrm{a}}$	$109.0\pm5.0^{\rm a}$
	Upland	$102.5\pm2.7^{\rm a}$	109.0 ± 4.6^a	$106.9 \pm 4.2^{\mathrm{a}}$	109.6 ± 1.8^{a}
LAR $(m^2 kg^{-1})$	Coastal	$4.13\pm0.37^{\text{b}}$	5.64 ± 0.11^{a}	4.09 ± 0.35^{b}	4.98 ± 0.42^{ab}
	Upland	4.54 ± 0.21^{ab}	5.43 ± 0.28^{ab}	4.73 ± 0.37^{ab}	5.42 ± 0.18^{ab}
Leaf mass fraction (%)	Coastal	39.6 ± 2.7^{e}	52.1 ± 1.8^{abc}	40.6 ± 2.5^{de}	52.5 ± 2.2^{abc}
	Upland	46.2 ± 1.8^{cde}	58.3 ± 1.7^{ab}	49.4 ± 2.6^{bcd}	$59.3\pm1.6^{\rm a}$
Stem mass fraction (%)	Coastal	11.6 ± 0.9^{cd}	17.2 ± 0.9^{a}	$10.9\pm0.8^{\rm d}$	15.0 ± 0.7^{abc}
	Upland	13.7 ± 0.6^{abcd}	15.4 ± 0.9^{ab}	13.5 ± 0.6^{bcd}	15.7 ± 0.9^{ab}
Tuber mass fraction (%)	Coastal	17.2 ± 2.5^{a}	3.7 ± 0.5^{cd}	13.8 ± 2.3^{ab}	3.8 ± 0.6^{cd}

Table 2-3 (continued)		Treatment	(m ² Kg ⁻¹)
Parameter	Genotype	$C_{\rm A}T_{\rm A}$	$C_{\rm A}T_{\rm E}$
	Upland	8.6 ± 1.2^{ab}	3.3 ± 0.5^{cd}
Root mass fraction (%)	Coastal	31.5 ± 2.4^{ab}	27.0 ± 1.6^{abc}
	Upland	31.5 ± 2.1^{ab}	23.0 ± 1.8^{bc}
Root/Shoot ratio	Coastal	$1.06\pm0.18^{\rm a}$	0.45 ± 0.03^{bc}
	Upland	0.69 ± 0.06^{ab}	0.36 ± 0.03^{c}
Leaf gas exchange			
$A_{\rm sat}$ (µmol m ⁻² s ⁻¹)	Coastal	9.6 ± 1.1^{ab}	$8.8\pm0.6^{\rm b}$
	Upland	10.1 ± 0.7^{ab}	9.4 ± 0.9^{ab}
$g_{\rm s} ({\rm mol}\;{\rm m}^{-2}\;{\rm s}^{-1})$	Coastal	$0.17\pm0.02^{\rm a}$	$0.17\pm0.02^{\rm a}$
	Upland	$0.19\pm0.01^{\rm a}$	0.18 ± 0.02^{a}
$A_{\rm max} \ (\mu {\rm mol} \ {\rm m}^{-2} {\rm s}^{-1})$	Coastal	$19.6 \pm 1.0^{\mathrm{a}}$	18.8 ± 0.9^{ab}
	Upland	18.7 ± 1.2^{abc}	$19.8\pm1.2^{\rm a}$
$V_{\rm cmax} \ (\mu { m mol} \ { m m}^{-2} \ { m s}^{-1})$	Coastal	$60.6\pm5.9^{\rm a}$	52.5 ± 4.2^{ab}
	Upland	56.8 ± 3.5^{ab}	50.7 ± 3.4^{ab}
$J_{\rm max} (\mu { m mol} \ { m m}^{-2} \ { m s}^{-1})$ 4.0 ^{abcd}	Coastal	81.5 ± 4.3^{a}	66.7 ±
	Upland	76.7 ± 4.4^{ab}	69.4 ± 4.0^{abcd}
$J_{\rm max}/V_{\rm cmax}$	Coastal	1.37 ± 0.07^{ab}	1.28 ± 0.04^{b}
	Upland	1.35 ± 0.02^{ab}	1.38 ± 0.06^{ab}

 $C_{\rm E}T_{\rm A}$ $C_{\rm E}T_{\rm E}$



fraction (%)

Table 2-3 (continued)		Treatment	nol m ⁻² s ⁻¹)		
Parameter	Genotype			$C_{\rm E}T_{\rm A}$	$C_{\rm E}T_{\rm E}$
AT_L $\widehat{\Box}_{s}$			- ,		
Parameter A	Coastal	-0.019 ± 0.002^{a}	$-0.019 \pm 0.001^{a_2^3}$	$-0.022\pm0.002^{\text{a}}$	-0.021 ± 0.001^{a}
(hmo	Upland	-0.019 ± 0.001^{a}	$-0.023 \pm 0.004^{a_{2}}$	-0.020 ± 0.002^{a}	-0.023 ± 0.004^{a}
Parameter B	Coastal	$0.98\pm0.09^{\mathrm{a}}$	$0.98\pm0.07^{\rm a}$	1.29 ± 0.12^{a}	1.21 ± 0.07^{a}
	Upland	$0.96\pm0.05^{\mathrm{a}}$	$1.25\pm0.20^{\mathrm{a}}$	1.12 ± 0.11^{a}	1.29 ± 0.22^a
Parameter C	Coastal	-4.04 ± 0.91^{a}	-4.20 ± 0.75^a	-7.71 ± 1.62^{a}	-6.27 ± 0.40^a
	Upland	-4.32 ± 0.15^{a}	-7.91 ± 1.90^{a}	-4.97 ± 1.31^{a}	-8.03 ± 1.94^a
$T_{\rm opt}(^{\circ}{\rm C})$	Coastal	25.8 ± 0.5^{a}	26.1 ± 0.6^a	$28.7\pm0.5^{\rm a}$	$28.0\pm0.8^{\rm a}$
	Upland	$25.3\pm0.1^{\text{a}}$	27.4 ± 1.1^{a}	28.2 ± 1.2^{a}	$28.4\pm0.6^{\rm a}$
A_{opt} (µmol m-2 s-1)	Coastal	8.6 ± 0.6^{a}	$8.6\pm0.6^{\rm a}$	$10.9\pm0.7^{\rm a}$	$10.7\pm0.8^{\rm a}$
	Upland	$7.9\pm0.7^{\rm a}$	8.9 ± 0.6^{a}	$10.9\pm0.4^{\rm a}$	$10.1\pm1.0^{\rm a}$
Carbohydrates					
Whole-plant St (mg g ⁻¹)	Coastal	16.2 ± 4.0^{ab}	$10.1 \pm 1.4^{\text{bc}}$	25.7 ± 3.7^{a}	$11.5 \pm 1.1^{\mathrm{abc}}$
	Upland	15.8 ± 4.3^{abc}	6.4 ± 0.9^{c}	20.5 ± 5.0^{ab}	8.9 ± 1.0^{bc}
Whole-plant Ss (mg g ⁻¹)	Coastal	37.2 ± 2.3^{a}	43.1 ± 2.5^{a}	$40.4\pm3.5^{\rm a}$	$46.2\pm1.5^{\rm a}$
	Upland	$35.8\pm0.4^{\rm a}$	$35.4 \pm 1.8^{\rm a}$	36.7 ± 3.3^{a}	$37.6\pm2.2^{\rm a}$
Whole-plant NSC (mg g ⁻¹) Coastal	53.4 ± 4.7^{ab}	53.3 ± 3.1^{ab}	66.1 ± 6.9^{a}	57.7 ± 1.9^{ab}
	Upland	51.6 ± 4.1^{ab}	$41.8\pm2.6^{\text{b}}$	57.2 ± 7.4^{ab}	46.5 ± 3.0^{ab}

The four [CO₂] and temperature treatments are: $C_A T_A$ (400 µl Γ^1 , 26 °C), $C_A T_E$ (400 µl Γ^1 , 30 °C), $C_E T_A$ (640 µl Γ^1 , 26 °C) and $C_E T_E$ (640 µl Γ^1 , 30 °C). DM, dry mass; LMA, leaf area per mass; LAR, leaf area ration; St, starch; Ss, soluble sugars; NSC, non-structural carbohydrates. Values represent means ± 1 SE (n = 10 for growth parameters and n = 5 for the others). Within each parameter, different superscript letters indicate means that are significantly different at P < 0.05 based on Tukey's pair-wise comparisons.

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Figure Error! No text of specified style in document.-1 Whole-plant dry mass (a), leaf area (b), leaf mass per area (LMA) (c), and leaf area ratio (LAR) (d) of *Telopea speciosissima* Coastal and Upland genotypes grown under the four [CO₂] and temperature treatments: $C_A T_A$ (400 µl l⁻¹, 26 °C; open blue), $C_A T_E$ (400 µl l⁻¹, 30 °C; open red), $C_E T_A$ (640 µl l⁻¹, 26 °C; closed blue), and $C_E T_E$ (640 µl l⁻¹, 30 °C; closed red). Values represent means ± 1 SE (*n* = 10).





Figure Error! No text of specified style in document.-2 Plant dry mass allocation of *Telopea speciosissima* Coastal and Upland genotypes grown under the four $[CO_2]$ and temperature treatments, including leaf mass fraction (a), stem mass fraction (b), tuber mass fraction (c), root mass fraction (d), and the Root/Shoot ratio (e). Values represent means ± 1 SE (n = 10).





Figure Error! No text of specified style in document.-**3** Light-saturated photosynthesis (A_{sat}) (a), stomatal conductance (g_s) (b), CO₂- and light-saturated assimilation rates (A_{max}) (c), maximum rate of photosynthetic carboxylation (V_{cmax}) (d), maximum rate of photosynthetic electron transport (J_{max}) (e), and the J_{max}/V_{cmax} ratio (f) of *Telopea speciosissima* Coastal and Upland genotypes grown under the four [CO₂] and temperature treatments. Values represent means ± 1 SE (n = 5).



12	(a)	Coas	stal ge 28.7 °C	notyp	e	
9		23 26.1	8.0 °C °C			
		25.8 °	°C			
6	~					
3	C _A C _A C _E	Τ _Α Τ _Ε Γ _Α				
0	$C_{\rm E}$	Ι _Ε				
12	(b)	Upla 2	and ge 8.2 °C	notyp	e	
9		2 27.	8.4 °C 4 °C			
6		25.4 °	С			
3						
0						
	15 20) 25	30	35	40	45
	L	eaf tem	perat	ure (°C)	

Figure Error! No text of specified style in document.-4 The simulated responses of CO₂ assimilation rates to leaf temperature (A/T_L) in *Telopea speciosissima* Coastal genotype (a) and Upland genotype (b) grown under the four [CO₂] and temperature treatments: $C_A T_A$ (dashed blue), $C_A T_E$ (dashed red), $C_E T_A$ (solid blue), and $C_E T_E$ (solid red). Curves represent the output of the averaged polynomial fits ($A_{sat} = A^*T_L^2 + B^*T_L + C$, where T_L is leaf temperature and A, B and C are the fitted parameters shown in Table 2-3) from 4 seedlings for each genotype per treatment. Coloured triangles and texts around the top of simulated curves indicate the photosynthetic thermal optimums (T_{opt}) under different treatments.

Non-structural carbohydrates (NSC)

Concentrations of whole-plant starch (St), soluble sugars (Ss), and non-structural carbohydrates (NSC) all varied between genotypes, but only St and NSC were significantly influenced by growth [CO₂] and temperature (Fig. 2-5; Table 2-2 and 2-3). Across treatments, the Coastal genotype had *c*. 25% and 15% higher whole-plant St and Ss, respectively, resulting in 18% higher (on average) NSC compared with the Upland genotype. $C_{\rm E}$ stimulated whole-plant St by 35%, while $T_{\rm E}$ reduced whole-plant St by 52%. No significant [CO₂] or temperature effect was found on whole-plant Ss. Consequently, $C_{\rm E}$ increased whole-plant NSC by 14%, but $T_{\rm E}$ decreased whole-plant NSC by 13% (Fig. 2-5c).

Across $[CO_2]$ and temperature treatments, the Coastal genotype had higher stem, tuber and root St, but similar leaf St when compared with the Upland genotype (Tables 2-2 and 2-4). Regardless of genotype, C_E stimulated leaf St by c. 65% but did not change St in other organs. T_E decreased leaf, stem and tuber St by 65%, 54% and 52%, respectively, without affecting root St. Averaged across treatments, the Coastal genotype had 25% higher leaf Ss and 10% higher root Ss, compared with the Upland genotype (Tables 2-2 and 2-4). C_E reduced stem Ss by 13%, but increased tuber and root Ss by 24% and 35%, respectively. T_E decreased root Ss for both genotypes, but reduced tuber Ss for the Upland genotype only (significant genotype × temperature interaction). For the Upland genotype, the positive effect of C_E on root Ss was offset by T_E (significant $[CO_2] \times$ temperature interaction), resulting in a significant genotype $\times [CO_2] \times$ temperature interaction (Tables 2-2 and 2-4).

40	(a)	$C_A T_A$	$C_E T_A$
		$C_A T_E$	$C_E T_E$
30			
20			
10			
10			
0			
60	ſb		
00)		
45			
30			
15			
0			
80			
	(c)		
60			
40			
20			
20			
0			
0	Coastal	Upl	and
	Gen	otype	

Figure Error! No text of specified style in document.-5 Whole-plant starch (a), soluble sugars (b), and nonstructural carbohydrates (NSC) (c) of *Telopea speciosissima* Coastal and Upland genotypes grown under the four [CO₂] and temperature treatments. Values represent means ± 1 SE (n = 5).

Table Error! No text of specified style in document.-4 Summary of means for starch and soluble sugar concentrations in different organs $\overline{\mathbf{x}}$ (leaf, stem, tuber and root) of *Telopea speciosissima* Coastal and Upland genotypes grown under the four [CO₂] and temperature $\overline{\mathbf{x}}$ $\overline{\mathbf{x}$

		Treatment			
Parameter	Genotype	$C_{\rm A}T_{\rm A}$	$C_{\rm A}T_{\rm E}$	$C_{\rm E}T_{\rm A}$	$C_{\rm E}T_{\rm E}$
Starch					
Leaf (mg g^{-1})	Coastal	19.4 ± 4.4^{abc}	9.2 ± 2.4^{cd}	41.0 ± 7.4^{a}	11.3 ± 0.6^{bcd}
	Upland	25.3 ± 7.6^{abc}	6.3 ± 0.8^{d}	30.8 ± 6.9^{ab}	12.4 ± 2.1^{bcd}
Stem (mg g^{-1})	Coastal	6.2 ± 1.2^{a}	2.6 ± 0.6^{abc}	$6.5\pm1.2^{\rm a}$	4.1 ± 1.5^{abc}
	Upland	4.3 ± 0.5^{ab}	2.0 ± 0.6^{bc}	3.7 ± 0.7^{abc}	$1.3 \pm 0.1^{\circ}$
Tuber (mg g^{-1})	Coastal	16.8 ± 4.8^{a}	7.2 ± 2.6^{ab}	$16.9\pm5.7^{\rm a}$	5.3 ± 1.5^{ab}
	Upland	2.8 ± 0.5^{ab}	1.9 ± 0.7^{b}	6.3 ± 1.1^{ab}	3.1 ± 0.1^{ab}
Root (mg g^{-1})	Coastal	$15.6\pm6.4^{\rm a}$	$17.1\pm1.7^{\rm a}$	10.9 ± 3.3^{a}	15.9 ± 2.2^{a}
	Upland	7.3 ± 1.7 a	11.5 ± 2.2^{a}	$10.8\pm2.4^{\rm a}$	7.4 ± 1.1^{a}
Soluble sugar					
Leaf (mg g^{-1})	Coastal	61.6 ± 4.0^{ab}	65.9 ± 3.4^{ab}	63.5 ± 5.3^{ab}	70.7 ± 4.1^{a}
	Upland	57.9 ± 2.2^{ab}	48.1 ± 2.9^{b}	49.0 ± 4.8^{b}	54.4 ± 4.1^{ab}
Stem (mg g^{-1})	Coastal	$22.8\pm3.3^{\rm a}$	$20.6\pm1.7^{\rm a}$	15.8 ± 0.9^{a}	20.9 ± 1.3^{a}

Table 2-4 (continued)

		Treatment				
Parameter	Genotype	$C_{\rm A}T_{\rm A}$	$C_{\rm A}T_{\rm E}$	$C_{\rm E}T_{\rm A}$	$C_{\rm E}T_{\rm E}$	
	Upland	22.8 ± 3.1^{a}	22.2 ± 0.9^a	$18.8 \pm 1.8^{\mathrm{a}}$	21.3 ± 1.1^a	
Tuber (mg g^{-1})	Coastal	$24.0\pm1.0^{\text{b}}$	26.7 ± 1.2^{ab}	31.1 ± 1.6^{ab}	32.7 ± 2.1^{ab}	
	Upland	25.3 ± 1.8^{ab}	22.8 ± 1.8^{b}	$35.5\pm4.7^{\mathrm{a}}$	23.6 ± 3.0^{b}	
Root (mg g^{-1})	Coastal	16.8 ± 0.9^{abc}	13.6 ± 0.3^{bc}	20.0 ± 1.6^{ab}	17.3 ± 1.7^{abc}	
	Upland	$12.4 \pm 0.8^{\circ}$	12.7 ± 0.7^{c}	24.2 ± 3.8^{a}	12.5 ± 1.1^{c}	

The four $[CO_2]$ and temperature treatments are: $C_A T_A$ (400 µl l⁻¹, 26 °C), $C_A T_E$ (400 µl l⁻¹, 30 °C), $C_E T_A$ (640 µl l⁻¹, 26 °C) and $C_E T_E$ (640 µl l⁻¹, 30 °C). Values represent means ± 1 SE (n = 5). Within each parameter, different superscript letters indicate means that are significantly different at P < 0.05 based on Tukey's pair-wise comparisons.
Relationships between biomass and physiological parameters

To assess those factors that may have regulated plant biomass, I examined the relationships between whole-plant dry mass and physiological parameters (i.e., photosynthetic traits and carbohydrate variables). Whole-plant dry mass increased with increasing A_{sat} , whole-plant Ss and leaf Ss, but decreased with increasing stem Ss (P < 0.05 in all cases), without significant differences between within-treatment correlations. No other associations between whole-plant dry mass and physiological traits were observed. A_{sat} accounted for only 10% of the variation in whole-plant dry mass (Fig. 2-6a), while whole-plant, leaf and stem Ss accounted for 20%, 22% and 11% of the variation in whole-plant dry mass, respectively (Fig. 2-6b,



(a) $r^2 = 0.10$ $r^2 = 0.20$ P < 0.01**(b)** P < 0.05 $A_{\rm sat}$ (µmol m⁻² s⁻¹) Whole-plant Ss (mg g^{-1}) 0 (c) $r^2 = 0.22$ $r^2 = 0.11$ (**d**) P < 0.05P < 0.01Leaf Ss (mg g^{-1}) Stem Ss (mg g^{-1})

Figure **Error! No text of specified style in document.**-6 The relationships between whole-plant dry mass and A_{sat} (a), whole-plant soluble sugars (whole-plant Ss) (b), leaf soluble sugars (leaf Ss) (c) and stem soluble sugars (stem Ss) (d) of *Telopea speciosissima* Coastal (circles) and Upland (triangles) genotypes grown under the four [CO₂] and temperature treatments: $C_A T_A$ (open blue), $C_A T_E$ (open red), $C_E T_A$ (closed blue) and $C_E T_E$ (closed red). There were five replicates per treatment, and each data point represents a single observation. Data were fitted using a linear regression (solid line). Data points for A_{sat} and stem Ss were log-log transformed before fitting. The adjusted r² value and its significance for each fitting are shown



PART 2: Drought experiment

At the onset of the *first drought*, growth parameters (i.e., stem length, basal diameter, and stem volume) and physiological traits (i.e., A_{sat} and g_s) were all similar between seedlings assigned to the *well-watered* and *drought* treatments within each [CO₂] and temperature treatment combination of each genotype ($P \ge 0.15$ in all cases), suggesting no bias in the initial allocation of seedlings to different watering treatments.

Plant growth

Across [CO₂], temperature and watering treatments, the Coastal genotype had higher whole-plant dry mass and leaf area than the Upland genotype (Fig. 3-1; Table 3-1). Regardless of watering treatment, C_E had positive effects on growth performance for both genotypes, while the positive growth response to T_E was only found in the Coastal genotype (significant genotype × temperature interaction; Table 3-1). The *drought* treatment, on the other hand, negatively affected dry mass accumulation and leaf growth in both genotypes, causing a reduction of 15–39% in whole-plant dry mass and an 18–43% decline in leaf area, respectively. However, declines induced by *drought* did not vary between genotypes or show significant difference among [CO₂] and temperature treatment combinations, indicating no interaction between *drought* and other treatments (Fig. 3-1; Table 3-1).

Stem volume in both genotypes had a similar pattern with plant dry mass in the response to experimental treatments, showing substantial increase under C_E but significant decline in the *drought*, as well as differentiated responses to T_E (Fig. 3-2; Table 3-2). Although a significant decrease in stem volume was found in *drought* seedlings (averaged across the five stages; P = 0.015) in relative to their *well-watered* counterparts, the effect of *drought* on stem volume was not significant until the *final harvest* (Fig. 3-2). For the other four stages (i.e., the *pre drought*, *first drought*, *recovery*, and *second drought*), there was no significant difference in stem volume between the two watering treatments. Changes in stem volume



 $A_{\rm sat}$ (µmol m⁻² s⁻¹)

32

induced by drought at the final harvest were mainly attributed to declines in the main stem basal diameter,



but not due to changes in the main stem length (Fig. A-1 and A-2; Table 3-2).

 $g_{\rm s} \,({
m mol} \,{
m m}^{-2}\,{
m s}^{-1})$

Table Error! No text of specified style in document.-5 Summary (P values) of four-way ANOVAs testing for the main and interactive effects of $[CO_2]$ (C), temperature (T) and watering (W) treatments on growth and carbohydrate parameters of two *Telopea speciosissima* genotypes (G)

	Growth	Carbohydrates					
Effect	Dry mass	Leaf ar ea	St	Ss	NSC		
G	< 0.001	< 0.001	0.068	0.404	0.196		
С	< 0.001	< 0.001	0.005	0.059	0.013		
Т	0.012	< 0.001	0.001	0.850	0.048		
W	< 0.001	< 0.001	< 0.001	< 0.001	0.667		
$\mathbf{G} imes \mathbf{C}$	0.662	0.383	0.535	0.527	0.837		
$\boldsymbol{G}\times\boldsymbol{T}$	0.009	0.005	0.608	0.276	0.400		
$\mathbf{C} imes \mathbf{T}$	0.558	0.975	0.557	0.163	0.422		
$\boldsymbol{G}\times \boldsymbol{W}$	0.503	0.785	0.018	0.049	0.026		
$\mathbf{C}\times\mathbf{W}$	0.561	0.984	0.087	0.549	0.761		
$\mathbf{T}\times\mathbf{W}$	0.497	0.970	< 0.001	0.081	0.280		
$G\times C\times T$	0.554	0.694	0.380	0.160	0.158		
$G\times C\times W$	0.493	0.457	0.719	0.274	0.318		
$G\times T\times W$	0.442	0.344	0.450	0.694	0.731		
$C\times T\times W$	0.513	0.636	0.083	0.208	0.085		
$G \times C \times T \times W$	0.696	0.340	0.435	0.223	0.513		

St, starch; Ss, soluble sugars; NSC, non-structural carbohydrates. Significant values (P < 0.05) are shown in bold. Analyses were run on data obtained from harvest samples, with ten replicates (n = 10) for growth and five replicates (n = 5) for carbohydrates.



Figure Error! No text of specified style in document.-7 Whole-plant dry mass (a and b) and leaf area (c and d) of *Telopea speciosissima* Coastal (the left panel) and Upland (the right panel) genotypes in *well-watered* and *drought* conditions exposed to four [CO₂] and temperature treatment combinations: $C_A T_A$ (400 µl l⁻¹, 26 °C; open blue), $C_A T_E$ (400 µl l⁻¹, 30 °C; open red), $C_E T_A$ (640 µl l⁻¹, 26 °C; closed blue), and $C_E T_E$ (640 µl l⁻¹, 30 °C; closed red). Values represent means ± 1 SE (n = 10).



Table Error! No text of specified style in document.-6 Summary (P values) of four-way repeated measures ANOVAs testing for the main and interactive effects of $[CO_2]$ (C), temperature (T) and watering (W) treatments on growth and gas exchange parameters of two *Telopea speciosissima* genotypes (G)

	Growth			Gas excha	inge
Effect	Length	Diameter	Volume	$A_{\rm sat}$	$g_{ m s}$
G	< 0.001	0.179	< 0.001	0.765	0.558
С	0.002	0.002	< 0.001	< 0.001	0.732
Т	< 0.001	0.045	< 0.001	< 0.001	< 0.001
W	0.821	0.030	0.015	< 0.001	< 0.001
$\mathbf{G} \times \mathbf{C}$	0.419	0.756	0.804	0.950	0.300
$\boldsymbol{G}\times\boldsymbol{T}$	< 0.001	0.021	< 0.001	0.823	0.938
$\mathbf{C} imes \mathbf{T}$	0.599	0.472	0.369	0.292	< 0.001
$\boldsymbol{G}\times \boldsymbol{W}$	0.959	0.348	0.268	0.264	0.013
$-C \times W$	0.876	0.845	0.663	0.003	0.687—
$\mathbf{T}\times\mathbf{W}$	0.484	0.475	0.884	0.014	0.376
$\overline{G} \times C \times T$	0.091	0.844	0.938	0.714	0.251
$G\times C\times W$	0.724	0.665	0.855	0.597	0.612
$G\times T\times W$	0.769	0.976	0.519	0.224	0.486
$C\times T\times W$	0.771	0.139	0.680	0.300	0.353
$G\times C\times T\times W$	0.458	0.780	0.894	0.592	0.056

 A_{sat} , light-saturated photosynthesis; g_s , stomatal conductance. Significant values (P < 0.05) are shown in bold. Analyses were run on data obtained during the experiment (multiple measurements), with ten _____ replicates (n = 10) for growth parameters and five replicates (n = 5) for gas exchange traits.



Figure Error! No text of specified style in document.-8 Progression of stem volume in *Telopea* speciosissima Coastal (the left panel) and Upland (the right panel) genotypes in well-watered (closed symbols) and *drought* (open symbols) conditions subjected to *ambient* (T_A ; blue) and *elevated* (T_E ; red) temperatures and *ambient* (C_A ; the top panel) and *elevated* (C_E ; the bottom panel) [CO₂] during the experimental stages: *pre drought* (Stage Pre), *first drought* (Stage D1), *recovery* (Stage R), *second drought* (Stage D2), and *final harvest* (Stage H). Values represent means ± 1 SE (n = 10).

Non-structural carbohydrates

The *drought* treatment had contrasting effects on whole-plant St and Ss, and the effect size on each parameter varied between genotypes (significant genotype × watering interactions) (Fig. 3-3; Table 3-1). Averaged across genotypes, $[CO_2]$ and temperature treatments, *drought* seedlings decreased whole-plant St by 56% but increased whole-plant Ss by 23%, when compared with *well-watered* seedlings. The negative effect of *drought* on whole-plant St was larger in the Coastal genotype (-61%) than in the Upland genotype (-49%). In contrast, the Coastal genotype showed a smaller increase (+12%) in whole-plant Ss under *drought* conditions, when compared with the increase in the Upland genotype (+35%). As a consequence of the opposite effects and the different effect sizes of *drought* on St and Ss, the *drought* treatment diminished the difference in whole-plant NSC between genotypes (significant genotype × watering interaction; Table 3-1). In addition, for both genotypes, the negative effect of *drought* on whole-plant St differed between temperature treatments (significant temperature × watering interaction; Fig. 3-3; Table 3-1). Averaged across genotypes and [CO₂] treatments, a larger decline in whole-plant St induced by *drought* was observed in T_A (-71%), when compared with the decrease in T_E (-22%).

		Coastal genotype			Upland genotype			
40	(a)		$C_A T_A$	C _E T _A	(b)			40
30			C _A I _E	CELE				30
20								20
10								10
0								0
60 (d)	(c)							60
45								45
30								30
15								15
0								0
8 0	(e)				(f			80
60								60
40								40
20								20
0	Well-	-watered	Drou	ght	Well-	watered	Drought	0

Figure Error! No text of specified style in document.-9 Whole-plant starch (a and b), soluble sugars (c and d), and non-structural carbohydrates (NSC) (e and f) of *Telopea speciosissima* Coastal and Upland genotypes in *well-watered* and *drought* conditions exposed to four $[CO_2]$ and temperature treatment combinations. Values represent means ± 1 SE (n = 5). Other details are as described for Fig. 3-1.



Leaf gas exchange

During the experimental period, both photosynthetic rates (A_{sat}) and stomatal conductance (g_s) did not differ between the two genotypes, but were significantly affected by temperature or watering treatments (Fig. 3-4 and 3-5; Table 3-2). C_E stimulated A_{sat} of both genotypes, but the magnitudes of stimulation varied between watering treatments. Averaged across stages, genotypes and temperature treatments, a larger increase of A_{sat} under C_E was found in *well-watered* seedlings (+40%) when compared with *drought* seedlings (+28%), suggesting a significant genotype × watering interaction. In contrast, T_E overall tended to decrease A_{sat} of both genotypes at both [CO₂] treatments, but the negative effect was only significant for the *well-watered* treatment (significant temperature × watering interaction), leading to an average 17% decline in A_{sat} of *well-watered* seedlings (Fig. 3-4). The *drought* treatment substantially decreased A_{sat} and g_s for both genotypes (Fig. 3-4 and 3-5; Table 3-2). The decline of g_s under *drought* was larger in the Coastal genotype (-52%; averaged across stages, [CO₂] and temperature treatments) than in the Upland genotype (- 39%), indicating a significant genotype × watering interaction. In addition, the decline in g_s induced by T_E was only significant under C_E (-31% on average across stages, genotypes and watering treatments; Fig. 3-5; Table 3-2).



Figure Error! No text of specified style in document.-**10** Progression of light-saturated photosynthesis (A_{sat}) in *Telopea speciosissima* Coastal (the left panel) and Upland (the right panel) genotypes in *well-watered* (closed symbols) and *drought* (open symbols) conditions subjected to *ambient* (T_A ; blue) and *elevated* (T_E ; red) temperatures and *ambient* (C_A ; the top panel) and *elevated* (C_E ; the bottom panel) [CO₂] during the four experimental stages: *pre drought* (Stage Pre), *first drought* (Stage D1), *recovery* (Stage R). Values represent

means ± 1 SE (n = 5).





Figure Error! No text of specified style in document.-11 Progression of stomatal conductance (g_s) in *Telopea speciosissima* Coastal and Upland genotypes in *well-watered* and *drought* conditions subjected to four [CO₂] and temperature treatment combinations during the four experimental stages. Values represent means ± 1 SE (n = 5). Other details are as described for Fig. 3-4.

The relationships between A_{sat} and soil VWC were fitted with three-parameter sigmoid functions. Overall, no significant difference between the two genotypes was found in any of the three fitted parameters at any [CO₂] and temperature treatment combination, suggesting no intraspecific variation in the sensitivity of A_{sat} to declines in soil water content (Fig. 3-6; Table 3-3). Regardless of temperature treatment, when soil water was not limiting, the estimated asymptote for A_{sat} was higher under C_E than C_A by 36% in the Coastal genotype and by 20% in the Upland genotype, respectively. However, the estimated asymptote for A_{sat} did not differ between temperature treatments for both genotypes (Fig. 3-6a and 6b; Table 3-3). As drought stress intensified (i.e., soil water content decreased), A_{sat} of both genotypes converged among the four [CO₂] and temperature treatment combinations, thereby promoting 50% loss of A_{sat} at similar soil water content across [CO₂] and temperature treatments. In other words, the inflection point (VWC_{mid}) of each sigmoid regression did not differ among [CO₂] and temperature treatments for both genotypes (Fig. 3-6a and 6b; Table 3-3). The relationships between g_s and soil VWC were assessed by linear regressions (on log-log scales). The linear fitting parameters did not differ between genotypes or among [CO₂] and temperature treatments, suggesting that there was no intraspecific variation in the sensitivity of g_s to declines in soil water content, and that the sensitivity was not affected by either [CO₂] or temperature (Fig. 3-6c and 6d; Table 3-4).



Figure Error! No text of specified style in document.-12 Light-saturated photosynthesis (A_{sat} ; a and b) and stomatal conductance (g_s ; c and d) of *Telopea speciosissima* Coastal and Upland genotypes in the *drought* treatment as a function of soil VWC exposed to *ambient* (T_A ; blue) and *elevated* (T_E ; red) temperatures and *ambient* (C_A ; circles) and *elevated* (C_E ; triangles) [CO₂]. Data are fitted for each of the four [CO₂] and temperature treatment combinations: $C_A T_A$ (the blue dash line), $C_A T_E$ (the red dash line), $C_E T_A$ (the blue solid line), and $C_E T_E$ (the red solid line). Data for A_{sat} are fitted with a three-parameter sigmoid regression, and data for g_s are fitted with a linear regression on log-log scales. Fit parameters are shown in Table 3-3 and Table 3-4, respectively.

Table Error! No text of specified style in document.-7 Summary of parameters in the fitted sigmoid regressions between light-saturated photosynthesis (A_{sat}) and soil VWC of *Telopea speciosissima* Coastal and Upland genotypes grown under the four [CO₂] and temperature treatments

Genotype	Treatment	t R ²	y asym		k		VWC _{mid}	
			Estimate	95% CI	Estimate	95% CI	Estimate	95% CI
Coastal	$C_{\rm A}T_{\rm A}$	0.768	12.254b	10.919,	189.748	46.822, 332.673	0.043	0.038, 0.048
13.589					49.938	-4.310, 104.187	0.041	0.021, 0.062
	$C_{\rm A}T_{\rm E}$	0.610	10.598b	7.977, 13.220	117.169	-2.738, 237.077	0.054	0.040, 0.067
	$C_{\rm E}T_{\rm A}$	0.921	17.340a	16.283, 18.396	104.177	16.884, 191.469	0.050	0.042, 0.058
	$\hat{C} \hat{T} = 0.8$ 16.103	330 1	3.315b 12	2.152, 14.479		34.022, 127.606	0.059	
Upland	A A				80.814			0.048, 0.069
	$C_{\rm A}T_{\rm E}$	0.854	12.225b	10.730, 13.721	165.537	-122.958, 454.032	0.057	0.042, 0.073
	$C_{\rm E}T_{\rm A}$	0.817	16.877a	15.521, <u>1</u> 8.233	74.611 ₋₁	26.040, 123.182	-1 0.062	0.052, 0.072
	<i>C</i> _E <i>T</i> _E 15.215	0.683	13.444b	11.672,	96.327	-45.675, 238.330	0.049	0.029, 0.069

The four [CO₂] and temperature treatments are: $C_A T_A$ (400 µl1 , 26 °C), $C_A T_E$ (400 µl1 , 30 °C), $C_E T_A$ (640 µl1 , 26 °C) and $C_E T_E$ (640 µl1⁻¹, 30 °C). The three-parameter sigmoid regressions were fitted as: $y = y_{asym} / (1 + e^{(-(VWC - VWCmid)/k)})$, where y_{asym} is the estimated asymptote for each regression, VWC_{mid} is the inflection point of soil VWC (where $y = y_{asym} / 2$) and *k* is a scaling parameter. Adjusted R² values (*P* < 0.001 in all cases) indicate the goodness-of-fit for regressions. Different letters indicate a significant difference among [CO₂] and temperature treatments for each parameter of each genotype based on the 95% confidence interval (i.e., 95% CI).



Table Error! No text of specified style in document.-8 Summary of parameters in the fitted linear regressions between stomatal conductance (g_s) and soil VWC of *Telopea speciosissima* Coastal and Upland genotypes grown under the four [CO₂] and temperature treatments

Genotype y ₀	Treatme	nt R ²	m				
			Estimate	e 95% CI	Estimate	95% CI	
Coastal	$C_{\rm A}T_{\rm A}$	0.600	0.609	0.084,	1.520	1.052, 1.988	
1.134					1.356	0.738, 1.974	
	$C_{\rm A}T_{\rm E}$	0.548	0.536	-0.255, 1.327	1.842	1.485, 2.199	
	$C_{-}T_{-}$	0 792	0 846	0 463 1 228	1.675 1.238	0.826, 1.650	
	$C_{\rm E}T_{\rm E}$	0.759	0.009	-0.237,	1.075		
	0.634						
Upland	A A					1.318, 2.032	
	$C_{\rm A}T_{\rm E}$	0.687	0.848	0.289, 1.407	1.729	1.229, 2.230	
	$C_{\rm E}T_{\rm A}$	0.833	0.833	0.524, 1.141	1.862	1.547, 2.178	
	$C_{\rm E}T_{\rm E}$	0.734	0.284	-0.060,	1.372	1.059, 1.684	
	0.628						

The four [CO₂] and temperature treatments are: $C_A T_A$ (400 µl 1 , 26 °C), $C_A T_E$ (400 µl 1 , 30 °C), $C_E T_A$ (640 µl 1⁻¹, 26 °C) and $C_E T_E$ (640 µl 1⁻¹, 30 °C). Linear regressions were fitted on log-log scales: $\log_{10}(y) = y_0 + m \times \log_{10}(VWC)$, where y_0 and *m* are the intercept and slope for each regression, respectively. Adjusted R² values (*P* < 0.001 in all cases) indicate the goodness-of-fit for regressions. 95% CI stands for the 95% confidence interval.



Resistance to drought

The capacity of *T. speciosissima* seedlings in resisting drought stress declined with plant size (i.e., the covariate; stem volume in this case), and there was a significant effect of temperature treatment on the capacity after removing the variance accounted for by the plant size (Fig. 3-7). Compared with T_A , increase in temperature negatively affected the capacity of seedling resistance to drought, leading to a quicker closure of stomata as drought progressed. At any common plant size, T_E accelerated the rates of g_s decline under *drought*, advancing the time for *drought* seedlings to become physiologically stressed by 1.5 weeks on average (Fig. 3-7). Apart from the temperature effect, the capacity of seedling resistance to drought did not vary between genotypes or $[CO_2]$ treatments, or among the treatment combinations.



Figure Error! No text of specified style in document.-13 Drought resistance (time for a seedling becoming physiologically stressed) versus plant size (stem volume) of drought-treated *Telopea speciosissima* exposed to *ambient* (T_A ; blue) and *elevated* (T_E ; red) temperatures and *ambient* (C_A ; circles) and *elevated* (C_E ; triangles) [CO₂]. Data are fitted with exponential regressions based on a generalized linear model (P < 0.001, $R^2 = 0.927$). Fittings for temperature treatments are shown in the plot.

Discussion

Intraspecific variation in woody plant responses to warming and

elevated [CO₂]

Significant intraspecific variation in growth plasticity between the two *T. speciosissima* genotypes was observed when responding to warming, with the Coastal genotype exhibiting greater increments in growth traits such as whole-plant dry mass, leaf area and SMF, compared to the Upland genotype. The differentiation between genotypes in growth response to warming reported here is consistent with the general prediction that plant populations may exhibit genetic variation in phenotypic plasticity (Donohue *et al.*, 2001; Alpert & Simms, 2002; Van Kleunen & Fischer, 2005; Aspinwall *et al.*, 2015). It has been

suggested that there is predictable intraspecific variation in the capacity of woody species to respond to $T_{\rm E}$ (Saxe *et al.*, 2001; Weston & Bauerle, 2007; Weston *et al.*, 2007; Way & Oren, 2010; Drake *et al.*, 2015). For example, Drake *et al.* (2015) studied 21 provenances of two widely distributed eucalyptus species (*Eucalyptus tereticornis* and *Eucalyptus grandis*) grown in conditions simulating ambient summer temperatures at seed origin and warmed temperatures (+ 3.5 °C), and found that the effect of warming on plant biomass and leaf area strongly interacted with the provenance's climate-of-origin. Similarly, I found that the growth capacity of woody plants in response to warming may vary among genotypes from contrasting climates.

Unlike other studies showing intraspecific variation in plasticity of physiological traits such as photosynthetic variables under $T_{\rm E}$ (Weston & Bauerle, 2007; Weston *et al.*, 2007; Drake *et al.*, 2015), differentiation in physiological plasticity of the two *T. speciosissima* genotypes in response to warming was largely absent in this study. $T_{\rm E}$ had similar effects on most physiological traits between the two genotypes, despite that there was significant genotype by temperature interaction on a few physiological traits including $g_{\rm s}$, $A/T_{\rm L}$ parameter C and tuber Ss. This phenomenon suggests that the effect of warming on growth plasticity was not parallel with the effect of warming on physiological plasticity in this study. This pattern may be attributed to the difference in plant size between the two *T. speciosissima* genotypes. Under

warming, both genotypes allocated more biomass to the above-ground for vegetative growth, as indicated by the reduced Root/Shoot ratio and the increased LAR, but the magnitudes of these changes did not differ between genotypes. However, the Coastal seedlings were bigger than the Upland counterparts, and therefore the Coastal genotype allocated more mass in essence to leaves and stems under warming conditions. In such circumstances, when compared with the Upland genotype, the Coastal genotype not only had greater whole-plant leaf area, but also showed greater increase in leaf area to warming, which is possibly the primary cause of the intraspecific variation in growth (i.e., biomass) response to temperature between the two *T. speciosissima* genotypes in this study.

Despite the fact that many traits (including growth and physiology) measured in this study showed a significant response to $C_{\rm E}$, no interaction between genotype and [CO₂] was found for any of the growth or physiological traits, indicating that the two *T. speciosissima* genotypes had similar phenotypic plasticity under $C_{\rm E}$. Although most studies on intraspecific variation in woody species responsiveness to $C_{\rm E}$ demonstrate substantial intraspecific differentiation in the responses of plant growth and/or physiology to changing [CO₂] (Ceulemans *et al.*, 1996; Dickson *et al.*, 1998; Isebrands *et al.*, 2001; Mohan *et al.*, 2004; Cseke *et al.*, 2009), some studies show limited intraspecific variation in woody plant responsiveness to $C_{\rm E}$ (e.g., Cantin *et al.*, 1997), In this study, both genotypes of *T. speciosissima* were equally limited by carbon availability and therefore showed strong increases in leaf area (29%) and mass production (35%) when grown in $C_{\rm E}$. Subsequently, rising [CO₂] is not likely to generate differential responses in genotypes of *T. speciosissima* in future climates.

I did not observe significant interaction between temperature and $[CO_2]$ in most traits measured in this study, except the ratio of J_{max}/V_{cmax} and the root Ss, suggesting that the effects of T_E and C_E were generally independent in the two *T. speciosissima* genotypes. There is no clear trend in the literature for the interactive effects of temperature and $[CO_2]$ on woody plant species. Many studies show that C_E is likely to interact with T_E , synergistically affecting woody plant growth and/or physiology (Callaway *et al.*, 1994; Peltola *et al.*, 2002; Ghannoum *et al.*, 2010a; Ayub *et al.*, 2011; Wang *et al.*, 2012). However, findings from the present study are consistent with other studies indicating that the effects of increasing $[CO_2]$ and warming are additive (Lewis *et al.*, 2001; Lloyd & Farquhar, 2008; Ghannoum *et al.*, 2010b; Lewis *et al.*, 2013). In addition, I also did not find significant interactive effects of temperature and $[CO_2]$ on the genotypic variation in phenotypic plasticity for most traits, except in root Ss. Especially for traits that showed interaction between genotype and temperature (i.e., plant dry mass, leaf area, SMF, g_s , A/T_L parameter C and tuber Ss), genotypic variation in phenotypic plasticity under T_E was not affected by changes in $[CO_2]$. I suggest that the lack of interactive effects of temperature and $[CO_2]$ on genotypic variation in phenotypic plasticity in this study may be partially due to the absence of interactive effects of temperature and $[CO_2]$ on plant growth and physiology.

Association between phenotypic plasticity and source environment variability of woody plant

populations

Plant populations usually show genetic differentiation in phenotypic plasticity and the divergence among populations may be influenced by the pattern of environmental variation. A long-standing hypothesis suggests that greater levels of environmental variability will select for genotypes with greater phenotypic plasticity (Galloway, 1995; Ackerly *et al.*, 2000; Weinig, 2000; Donohue *et al.*, 2001; Alpert & Simms, 2002; Gianoli & Gonzalez-Teuber, 2005; Van Kleunen & Fischer, 2005). Although testing this hypothesis on woody plant species is limited, there is at least one case study that supports the theory (Drake *et al.*, 2015). Specifically, this case study on two widespread eucalyptus species (*E. tereticornis* and *E. grandis*) showed that, for both species, provenances originating from cooler and more variable temperature climates exhibited higher plasticity in growth and photosynthetic capacity under warming, when compared with provenances from warmer and more uniform temperature climates (Drake *et al.*, 2015).

Results from this study contradict the current paradigm. I observed that the Coastal genotype of *T*. *speciosissima* (warmer and less variable temperature environments) rather than the Upland genotype (cooler and more variable temperature environments), exhibited higher growth plasticity in response to $T_{\rm E}$. The differentiation in phenotypic plasticity among plant populations may be associated with source environment



variability and linked to the intrinsic difference in adaptation to distinct source environments. Plant populations usually are highly adapted to local conditions, showing the greatest fitness in their home environments (Savolainen *et al.*, 2007; Hereford, 2009; Wang *et al.*, 2010). The upland region in this study is *c*. 2–5 °C cooler than the coastal region (Table 2-1), and the temperature difference between these regions has been estimated to be larger during the Last Glacial Maximum (Barrows *et al.*, 2001; Hesse *et al.*, 2003). This long-term temperature differential may have shaped and maintained the genetic differences between the coastal and upland populations of *T. speciosissima* (Rossetto *et al.*, 2011). The Upland genotype that might have been adapted to cooler temperatures, may not have the capacity to fully utilise warmer temperatures in terms of plant growth, and therefore showed lower growth plasticity in response to T_E when compared with the warmer-origin Coastal genotype. However, to more rationally explain why the results in this study contradict the long-standing paradigm, further studies with a more specific and thorough design (e.g., with both ecological and evolutionary aspects included) on *T. speciosissima* would be more informative.

3.4.1 No intraspecific variation in growth and photosynthetic responses to drought

Between the two *T. speciosissima* genotypes originating from contrasting environments with differentiated precipitation, differences in the declines of growth and photosynthesis induced by drought were largely absent across $[CO_2]$ and temperature treatments. In addition, their sensitivity of photosynthetic traits (i.e., A_{sat} and g_s) to declines in soil water content and the capacity of plant drought resistance (measured by the time for drought treated seedlings to become physiologically stressed) also did not differ between the two genotypes. These results collectively suggest that there is no intraspecific variation in the response to drought between the *T. speciosissima* genotypes in this study, contradicting observations on other woody species, in which plant populations from different precipitation regions usually showed differentiated responses under drought conditions (e.g., Ramirez-Valiente *et al.*, 2010; McLean *et al.*, 2014; Bansal *et al.*, 2015). Specifically, these studies indicated that woody plant populations originated from more mesic regions were usually more susceptible to drought (Cregg & Zhang, 2001; Silva *et al.*, 2006; Ramirez- Valiente *et al.*, 2010; Dutkowski & Potts, 2012; Robson *et al.*, 2012), while populations from more stressful

environments tended to be less responsive to water stress (Gratani *et al.*, 2003; Baquedano *et al.*, 2008; Aranda *et al.*, 2010; Bansal *et al.*, 2015).

The lack of intraspecific variation in response to drought in this study could be attributed to the fact that there might be no inherent difference in the capacity to cope with drought between the two *T*. *speciosissima* genotypes. Although the Coastal and the Upland genotypes were sampled from regions with different precipitation, both regions can be characterized as high rainfall regions (more than 850 mm per year) with no difference in precipitation variability (see Huang *et al.*, 2015), suggesting the relative uniformity of precipitation conditions between the two regions. Therefore, these two genotypes might have been adapted to somewhat similar non-water-stressed environments and may not differ in their inherent capacity of coping with water deficit. Similar results were found in a drought manipulating study on provenances of two widely distributed *Eucalyptus* species, where provenances originating from contrasting environments (tropical *vs.* temperate) did not show intraspecific variation in most growth and physiological responses to drought (Huang *et al.*, unpublished data). In that study, provenances were also selected from

regions with relatively sufficient precipitation (all > 890 mm rainfalls per year) and similar precipitation variability, despite that there was significant difference in the mean annual precipitation (MAP) between them. In contrast, woody plant populations exhibiting intraspecific variation in the drought responses usually distribute across low (MAP < 400 mm), mid (MAP between 400 and 800 mm) and high (MAP > 800 mm) rainfall regions (Aranda *et al.*, 2010; Ramirez-Valiente *et al.*, 2010; McLean *et al.*, 2014; Bansal *et al.*,

2015), or at least two contrasting rainfall regions (Cregg & Zhang, 2001; Gratani *et al.*, 2003; Silva *et al.*, 2006; Robson *et al.*, 2012), suggesting that these populations may possess inherent difference in their capacity to cope with water stress due to local adaptation.

3.4.2 Effects of $T_{\rm E}$ and $C_{\rm E}$ on woody plants in response to drought

I observed a significant effect of temperature on the capacity of seedling resistance to drought after removing the variance accounted for by the plant size. For both *T. speciosissima* genotypes, $T_{\rm E}$ accelerated


the rates of g_s decline under drought conditions and thereby reduced the time for drought seedlings to become physiologically stressed. Results from this study are consistent with the prevailing findings that an increase in air temperature usually exacerbates the negative impacts of water stress on woody plants (Adams *et al.*, 2009; Allen *et al.*, 2010; Duan *et al.*, 2013, 2014, 2015; Will *et al.*, 2013; Zhao *et al.*, 2013). However, the quicker closure of stomata under T_E did not reflect in the sensitivity of g_s as a function of soil VWC in this study. For both temperature treatments, g_s positively correlated with soil water content in a similar manner, suggesting that the nature of *T. speciosissima* stomata in response to declines in soil water content was not altered by changes in temperature. Therefore, I hypothesize that the negative impacts of T_E on the drought resistance of *T. speciosissima* seedlings may be working as follows: under drought conditions, higher temperatures will accelerate transpiration water loss through the increase in vapour pressure deficits, which will in turn speed up the drawdown of soil water content and hence create a positive feedback loop to magnify or exacerbate the negative effects of drought (De Boeck *et al.*, 2011; Will *et al.*, 2013; Teskey *et al.*, 2015).

By contrast, an increase in $[CO_2]$ neither impacted the capacity of plant drought resistance, nor altered the sensitivity of A_{sat} or g_s to declines in soil water content for both *T. speciosissima* genotypes in this study. In addition, C_E did not ameliorate the negative effects of T_E on drought resistance, suggesting that C_E may be a less strong determinant than T_E on regulating plant response to drought. Observations about the effects of C_E on woody plant drought response are considerably inconsistent in literature. Some studies indicate that C_E would lead to partial closure of stomata, thereby reducing transpiration water loss and mitigating the negative effects of drought on plant performance (Ambebe & Dang, 2010; Wertin *et al.*, 2010; Duan *et al.*, 2013; Lewis *et al.*, 2013); while other studies (Duan *et al.*, 2014, 2015) suggest that C_E may only have a negligible effect on woody plant response to drought, consistent with findings of this study. The absence of $[CO_2]$ effects on *T. speciosissima* drought response may be explained by the fact that g_s in this study overall did not differ between $[CO_2]$ treatments across all experimental stages, indicating that C_E did not significantly reduce g_s to improve plant water usage and therefore did not ameliorate the negative effects of drought. Although most woody plants show a significant decrease in g_s under C_E (Wullschleger *et al.*, 2002; Ainsworth & Long, 2005; Ainsworth & Rogers, 2007; Wang *et al.*, 2012), there are some



exceptions as well (Saxe *et al.*, 1998; Ellsworth, 1999; Lewis *et al.*, 2002; Ghannoum *et al.*, 2010a; Duan *et al.*, 2014, 2015). Given the inconsistence and complexity of $[CO_2]$ effects on plant drought response (Wullschleger *et al.*, 2002; Franks *et al.*, 2013), further studies with a systematic manner are necessary for exploring mechanisms that underpin woody plant response to drought and $C_{\rm E}$.

In conclusion, I found that the Coastal genotype of *T. speciosissima*, which originated from warmer and less variable temperature environments, showed greater plasticity in growth with warming than the Upland genotype from cooler and more variable temperature environments. On the other hand, $C_{\rm E}$ did not expose genotypic variation in growth or physiological responses, either individually or interactively with $T_{\rm F}$. These findings suggest that temperature will be more effective than [CO₂] in exposing intraspecific variation in growth plasticity for genetically differentiated woody plant populations under future climates. Overall, results from this study contradict the paradigm that genotypes from more variable climates will exhibit greater phenotypic plasticity in future climate regimes. The two T. speciosissima genotypes neither showed difference in their capacity in resisting to drought stress, nor exhibited differentiated declines in growth and photosynthesis under drought conditions across [CO₂] and temperature treatments, suggesting that there might be no inherent difference in their capacity to cope with drought. Regardless of genotype, $T_{\rm E}$ imposed a negative effect on plant drought resistance, accelerating the process of drought seedlings becoming physiologically stressed. In contrast, $C_{\rm E}$ did not affect the capacity of plant drought resistance or alter the sensitivity of photosynthesis to declines in soil water content for both T. speciosissima genotypes. Furthermore, $C_{\rm E}$ did not ameliorate the negative effects of $T_{\rm E}$ on drought response. Collectively, these findings suggest that woody plant populations originating from differentiated environments may not necessarily show intraspecific variation in response to drought under current climates or future climates. These results also indicate that temperature is likely to be stronger determinant than [CO₂] affecting the capacity of woody plants in resisting to drought in the context of climate change.



References

Ackerly DD, Dudley SA, Sultan SE, Schmitt J, Coleman JS, Linder CR, Sandquist DR, Geber MA, Evans AS, Dawson TE, Lachowicz MJ (2000) The evolution of plant ecophysiological traits: Recent advances and future directions. Bioscience, 50, 979-995.

Adams HD, Guardiola-Claramonte M, Barron-Gafford GA, Villegas JC, Breshears DD, Zou CB, Troch PA, Huxman TE (2009) Temperature sensitivity of drought-induced tree mortality portends increased regional die-off under global-change-type drought. Proceedings of the National Academy of Sciences, 106, 7063-7066.

Ahuja I, De Vos RCH, Bones AM, Hall RD (2010) Plant molecular stress responses face climate change. Trends in Plant Science, 15, 664-674.

Ainsworth EA, Long SP (2005) What have we learned from 15 years of free-air CO2 enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO2. New Phytologist, 165, 351-371.

Ainsworth EA, Rogers A (2007) The response of photosynthesis and stomatal conductance to rising [CO2]: mechanisms and environmental interactions. Plant, Cell & Environment, 30, 258-270.

Allen CD, Macalady AK, Chenchouni H, Bachelet D, Mcdowell N, Vennetier M, Kitzberger T, Rigling A, Breshears DD, Hogg EH, Gonzalez P, Fensham R, Zhang Z, Castro J, Demidova N, Lim JH, Allard G, Running SW, Semerci A, Cobb N (2010) A global overview of drought and heat-induced tree mortality reveals emerging climate change risks for forests. Forest Ecology and Management, 259, 660-684.

Alpert P, Simms EL (2002) The relative advantages of plasticity and fixity in different environments: when is it good for a plant to adjust? Evolutionary Ecology, 16, 285-297.

Ambebe TF, Dang QL (2010) Low moisture availability reduces the positive effect of increased soil temperature on biomass production of white birch (Betula papyrifera) seedlings in ambient and elevated carbon dioxide concentration. Nordic Journal of Botany, 28, 104-111.

Ameye M, Wertin TM, Bauweraerts I, Mcguire MA, Teskey RO, Steppe K (2012) The effect of induced heat waves on Pinus taeda and Quercus rubra seedlings in ambient and elevated CO2 atmospheres. New Phytologist, 196, 448-461.

Anderson JT, Panetta AM, Mitchell-Olds T (2012) Evolutionary and Ecological Responses to Anthropogenic Climate Change. Plant Physiology, 160, 1728-1740.

Aranda I, Alia R, Ortega U, Dantas AK, Majada J (2010) Intra-specific variability in biomass partitioning and carbon isotopic discrimination under moderate drought stress in seedlings from four Pinus pinaster populations. Tree Genetics & Genomes, 6, 169-178.

Aspinwall MJ, Loik ME, Resco De Dios V, Tjoelker MG, Payton PR, Tissue DT (2015) Utilizing intraspecific variation in phenotypic plasticity to bolster agricultural and forest productivity under climate change. Plant, Cell & Environment, 38, 1752-1764.

Atkin OK, Tjoelker MG (2003) Thermal acclimation and the dynamic response of plant respiration to temperature. Trends in Plant Science, 8, 343-351.

Ayub G, Smith RA, Tissue DT, Atkin OK (2011) Impacts of drought on leaf respiration in darkness and light in Eucalyptus saligna exposed to industrial-age atmospheric CO2 and growth temperature. New Phytologist, 190, 1003-1018.

Ballester J, Rodo X, Giorgi F (2010) Future changes in Central Europe heat waves expected to mostly follow summer mean warming. Climate Dynamics, 35, 1191-1205.

Bansal S, Harrington CA, Gould PJ, St Clair JB (2015) Climate-related genetic variation in drought-resistance of Douglas-fir (Pseudotsuga menziesii). Global Change Biology, 21, 947-958.

Banta JA, Ehrenreich IM, Gerard S, Chou L, Wilczek A, Schmitt J, Kover PX, Purugganan MD (2012) Climate envelope modelling reveals intraspecific relationships among flowering phenology, niche breadth and potential range size in Arabidopsis thaliana. Ecology Letters, 15, 769-777.

Baquedano FJ, Valladares F, Castillo FJ (2008) Phenotypic plasticity blurs ecotypic divergence in the response of Quercus coccifera and Pinus halepensis to water stress. European Journal of Forest Research, 127, 495-506.

Barnosky AD, Hadly EA, Bascompte J, Berlow EL, Brown JH, Fortelius M, Getz WM, Harte J, Hastings A, Marquet PA, Martinez ND, Mooers A, Roopnarine P, Vermeij G, Williams JW, Gillespie R, Kitzes J, Marshall C, Matzke N, Mindell DP, Revilla E, Smith AB (2012) Approaching a state shift in Earth's biosphere. Nature, 486, 52-58.

Barrows TT, Stone JO, Fifield LK, Cresswell RG (2001) Late Pleistocene glaciation of the Kosciuszko Massif, Snowy Mountains, Australia. Quaternary Research, 55, 179-189.

Bastos A, Gouveia C, Trigo R, Running S (2013) Comparing the impacts of 2003 and 2010 heatwaves in NPP over Europe. Biogeosciences Discussions, 10, 15879-15911.

Bates D, Maechler M, Bolker B, Walker S (2014) lme4: Linear mixed-effects models using Eigen and S4, R package version 1.1-7, http://CRAN.R-project.org/package=lme4.

Bauweraerts I, Ameye M, Wertin TM, Mcguire MA, Teskey RO, Steppe K (2014) Water availability is the decisive factor for the growth of two tree species in the occurrence of consecutive heat waves. Agricultural and Forest Meteorology, 189, 19-29.

Bauweraerts I, Wertin TM, Ameye M, Mcguire MA, Teskey RO, Steppe K (2013) The effect of heat waves, elevated [CO2] and low soil water availability on northern red oak (Quercus rubra L.) seedlings. Global Change Biology, 19, 517-528.

Bedon F, Villar E, Vincent D, Dupuy JW, Lomenech AM, Mabialangoma A, Chaumeil P, Barre A, Plomion C, Gion JM (2012) Proteomic plasticity of two Eucalyptus genotypes under contrasted water regimes in the field. Plant, Cell & Environment, 35, 790-805.

Beer C, Reichstein M, Tomelleri E, Ciais P, Jung M, Carvalhais N, Rodenbeck C, Arain MA, Baldocchi D, Bonan GB, Bondeau A, Cescatti A, Lasslop G, Lindroth A, Lomas M, Luyssaert S, Margolis H, Oleson KW, Roupsard O, Veenendaal E, Viovy N, Williams C, Woodward FI, Papale D (2010) Terrestrial Gross Carbon Dioxide Uptake: Global Distribution and Covariation with Climate. Science, 329, 834-838.

Berry J, Bjorkman O (1980) Photosynthetic Response and Adaptation to Temperature in Higher-Plants. Annual Review of Plant Physiology and Plant Molecular Biology, 31, 491-543.

Bobich EG, Barron-Gafford GA, Rascher KG, Murthy R (2010) Effects of drought and changes in vapour pressure deficit on water relations of Populus deltoides growing in ambient and elevated CO2. Tree Physiology, doi: 10.1093/treephys/tpq1036.

Bohnert HJ, Nelson DE, Jensen RG (1995) Adaptations to Environmental Stresses. The Plant Cell, 7, 1099-1111.

Bonan GB (2008) Forests and Climate Change: Forcings, Feedbacks, and the Climate Benefits of Forests. Science, 320, 1444-1449.

Boyer JS (1982) Plant Productivity and Environment. Science, 218, 443-448.

Bradshaw AD (1965) Evolutionary significance of phenotypic plasticity in plants. Advances in Genetics, 13, 115-155.

Bunn AG, Graumlich LJ, Urban DL (2005) Trends in twentieth-century tree growth at high elevations in the Sierra Nevada and White Mountains, USA. Holocene, 15, 481-488.

Callaway RM, Delucia EH, Thomas EM, Schlesinger WH (1994) Compensatory Responses of CO2 Exchange and Biomass Allocation and Their Effects on the Relative Growth-Rate of Ponderosa Pine in Different CO2 and Temperature Regimes. Oecologia, 98, 159-166.

Campbell C, Atkinson L, Zaragoza-Castells J, Lundmark M, Atkin O, Hurry V (2007) Acclimation of photosynthesis and respiration is asynchronous in response to changes in temperature regardless of plant functional group. New Phytologist, 176, 375-389.

Cantin D, Tremblay MF, Lechowicz MJ, Potvin C (1997) Effects of CO2 enrichment, elevated temperature, and nitrogen availability on the growth and gas exchange of different families of jack pine seedlings. Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere, 27, 510-520.

Carter KK (1996) Provenance tests as indicators of growth response to climate change in 10 north temperate tree species. Canadian Journal of Forest Research-Revue Canadianne De Recherche Forestiere, 26, 1089-1095.

Ceulemans R, Shao BY, Jiang XN, Kalina J (1996) First- and second-year aboveground growth and productivity of two Populus hybrids grown at ambient and elevated CO2. Tree Physiology, 16, 61-68.

Chaves MM (1991) Effects of Water Deficits on Carbon Assimilation. Journal of Experimental Botany, 42, 1-16.

Chaves MM, Maroco JP, Pereira JS (2003) Understanding plant responses to drought - from genes to the whole plant. Functional Plant Biology, 30, 239-264.

Chevin LM, Lande R, Mace GM (2010) Adaptation, Plasticity, and Extinction in a Changing Environment: Towards a Predictive Theory. PLoS Biology, 8, e1000357.

Ciais P, Reichstein M, Viovy N, Granier A, Ogee J, Allard V, Aubinet M, Buchmann N, Bernhofer C, Carrara A, Chevallier F, De Noblet N, Friend AD, Friedlingstein P, Grunwald T, Heinesch B, Keronen P, Knohl A, Krinner G, Loustau D, Manca G, Matteucci G, Miglietta F, Ourcival JM, Papale D, Pilegaard K, Rambal S, Seufert G, Soussana JF, Sanz MJ, Schulze ED, Vesala T, Valentini R (2005) Europe-wide reduction in primary productivity caused by the heat and drought in 2003. Nature, 437, 529-533.

Clark DA, Piper SC, Keeling CD, Clark DB (2003) Tropical rain forest tree growth and atmospheric carbon dynamics linked to interannual temperature variation during 1984-2000. Proceedings of the National Academy of Sciences of the United States of America, 100, 5852-5857.

Clark DB, Clark DA, Oberbauer SF (2010) Annual wood production in a tropical rain forest in NE Costa Rica linked to climatic variation but not to increasing CO2. Global Change Biology, 16, 747-759.

Collins M, Knutti R, Arblaster J, Dufresne J-L, Fichefet T, Friedlingstein P, Gao X, Gutowski WJ, Johns T, Krinner G, Shongwe M, Tebaldi C, Weaver AJ, Wehner M (2013) Long-term Climate Change: Projections, Commitments and Irreversibility. In: Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. (eds Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex V, Midgley PM) pp 1029-1136. Cambridge, United Kingdom and New York, NY, USA, Cambridge University Press.

Coumou D, Rahmstorf S (2012) A decade of weather extremes. Nature Climate Change, 2, 491-496.

Coumou D, Robinson A (2013) Historic and future increase in the global land area affected by monthly heat extremes. Environmental Research Letters, 8, 034018.

Coumou D, Robinson A, Rahmstorf S (2013) Global increase in record-breaking monthly-mean temperatures. Climatic Change, 118, 771-782.

Cramer W, Bondeau A, Woodward FI, Prentice IC, Betts RA, Brovkin V, Cox PM, Fisher V, Foley JA, Friend AD, Kucharik C, Lomas MR, Ramankutty N, Sitch S, Smith B, White A, Young-Molling C (2001) Global response of terrestrial ecosystem structure and function to CO2 and climate change: results from six dynamic global vegetation models. Global Change Biology, 7, 357-373.

Cregg BM, Zhang JW (2001) Physiology and morphology of Pinus sylvestris seedlings from diverse sources under cyclic drought stress. Forest Ecology and Management, 154, 131-139.

Cseke LJ, Tsai CJ, Rogers A, Nelsen MP, White HL, Karnosky DF, Podila GK (2009) Transcriptomic comparison in the leaves of two aspen genotypes having similar carbon assimilation rates but different partitioning patterns under elevated [CO2]. New Phytologist, 182, 891-911.

Cunningham SC, Read J (2006) Foliar temperature tolerance of temperate and tropical evergreen rain forest trees of Australia. Tree Physiology, 26, 1435-1443.

Dai A (2013) Increasing drought under global warming in observations and models. Nature Climate Change, 3, 52-58.

De Boeck HJ, Dreesen FE, Janssens IA, Nijs I (2010) Climatic characteristics of heat waves and their simulation in plant experiments. Global Change Biology, 16, 1992-2000.

De Boeck HJ, Dreesen FE, Janssens IA, Nijs I (2011) Whole-system responses of experimental plant communities to climate extremes imposed in different seasons. New Phytologist, 189, 806-817.

Della-Marta PM, Haylock MR, Luterbacher J, Wanner H (2007) Doubled length of western European summer heat waves since 1880. Journal of Geophysical Research: Atmospheres, 112, D15103, doi: 10.11029/12007JD008510.

Dickson RE, Coleman MD, Riemenschneider DE, Isebrands JG, Hogan GD, Karnosky DF (1998) Growth of five hybrid poplar genotypes exposed to interacting elevated CO2 and O3. Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere, 28, 1706-1716.

Donohue K, Pyle EH, Messiqua D, Heschel MS, Schmitt J (2001) Adaptive divergence in plasticity in natural populations of Impatiens capensis and its consequences for performance in novel habitats. Evolution, 55, 692-702.

Doughty CE, Goulden ML (2008) Are tropical forests near a high temperature threshold? Journal of Geophysical Research: Biogeosciences, 113, G00B07, doi:10.1029/2007JG000632.

Drake JE, Aspinwall MJ, Pfautsch S, Rymer PD, Reich PB, Smith RA, Crous KY, Tissue DT, Ghannoum O, Tjoelker MG (2015) The capacity to cope with climate warming declines from temperate to tropical latitudes in two widely distributed Eucalyptus species. Global Change Biology, 21, 459-472.

Dreesen FE, De Boeck HJ, Janssens IA, Nijs I (2012) Summer heat and drought extremes trigger unexpected changes in productivity of a temperate annual/biannual plant community. Environmental and Experimental Botany, 79, 21-30.

Duan H, Duursma RA, Huang G, Smith RA, Choat B, O'Grady AP, Tissue DT (2014) Elevated [CO2] does not ameliorate the negative effects of elevated temperature on drought-induced mortality in Eucalyptus radiata seedlings. Plant, Cell & Environment, 37, 1598-1613.

Duan H, O'Grady AP, Duursma RA, Choat B, Huang G, Smith RA, Jiang Y, Tissue DT (2015) Drought responses of two gymnosperm species with contrasting stomatal regulation strategies under elevated [CO2] and temperature. Tree Physiology, 35, 756-770.

Duan HL, Amthor JS, Duursma RA, O'Grady AP, Choat B, Tissue DT (2013) Carbon dynamics of eucalypt seedlings exposed to progressive drought in elevated [CO2] and elevated temperature. Tree Physiology, 33, 779-792.

Dutkowski GW, Potts BM (2012) Genetic variation in the susceptibility of Eucalyptus globulus to drought damage. Tree Genetics & Genomes, 8, 757-773.

Duursma RA, Barton CVM, Eamus D, Medlyn BE, Ellsworth DS, Forster MA, Tissue DT, Linder S, Mcmurtrie RE (2011) Rooting depth explains [CO2] × drought interaction in Eucalyptus saligna. Tree Physiology, 31, 922-931.

Ebell LF (1969) Variation in total soluble sugars of conifer tissues with method of analysis. Phytochemistry, 8, 227-233.

Ellsworth DS (1999) CO2 enrichment in a maturing pine forest: are CO2 exchange and water status in the canopy affected? Plant, Cell & Environment, 22, 461-472.

Fao (2006) Global forest resources assessment 2005: progress towards sustainable forest management. FAO Forestry Paper No. 147. Rome.

Feeley KJ, Wright SJ, Supardi MNN, Kassim AR, Davies SJ (2007) Decelerating growth in tropical forest trees. Ecology Letters, 10, 461-469.

Flexas J, Bota J, Escalona JM, Sampol B, Medrano H (2002) Effects of drought on photosynthesis in grapevines under field conditions: an evaluation of stomatal and mesophyll limitations. Functional Plant Biology, 29, 461-471.

Franks PJ, Adams MA, Amthor JS, Barbour MM, Berry JA, Ellsworth DS, Farquhar GD, Ghannoum O, Lloyd J, Mcdowell N, Norby RJ, Tissue DT, Von Caemmerer S (2013) Sensitivity of plants to changing atmospheric CO2 concentration: from the geological past to the next century. New Phytologist, 197, 1077-1094.

Frich P, Alexander LV, Della-Marta P, Gleason B, Haylock M, Tank AMGK, Peterson T (2002) Observed coherent changes in climatic extremes during the second half of the twentieth century. Climate Research, 19, 193-212.

Galloway LF (1995) Response to natural environmental heterogeneity: Maternal effects and selection on life-history characters and plasticities in Mimulus guttatus. Evolution, 49, 1095-1107.

Gauthier PP, Crous KY, Ayub G, Duan H, Weerasinghe LK, Ellsworth DS, Tjoelker MG, Evans JR, Tissue DT, Atkin OK (2014) Drought increases heat tolerance of leaf respiration in Eucalyptus globulus saplings grown under both ambient and elevated atmospheric [CO2] and temperature. Journal of Experimental Biology, 65, 6471-6485.

Ghannoum O, Phillips NG, Conroy JP, Smith RA, Attard RD, Woodfield R, Logan BA, Lewis JD, Tissue DT (2010a) Exposure to preindustrial, current and future atmospheric CO2 and temperature differentially affects growth and photosynthesis in Eucalyptus. Global Change Biology, 16, 303-319.

Ghannoum O, Phillips NG, Sears MA, Logan BA, Lewis JD, Conroy JP, Tissue DT (2010b) Photosynthetic responses of two eucalypts to industrial-age changes in atmospheric [CO2] and temperature. Plant, Cell & Environment, 33, 1671-1681.

Gianoli E, Gonzalez-Teuber M (2005) Environmental heterogeneity and population differentiation in plasticity to drought in Convolvulus chilensis (Convolvulaceae). Evolutionary Ecology, 19, 603-613.

Gratani L, Meneghini M, Pesoli P, Crescente MF (2003) Structural and functional plasticity of Quercus ilex seedlings of different provenances in Italy. Trees-Structure and Function, 17, 515-521.

Grime JP, Mackey JML (2002) The role of plasticity in resource capture by plants. Evolutionary Ecology, 16, 299-307.

Gunderson CA, Norby RJ, Wullschleger SD (2000) Acclimation of photosynthesis and respiration to simulated climatic warming in northern and southern populations of Acer saccharum: laboratory and field evidence. Tree Physiology, 20, 87-96.

Hacke UG, Jacobsen AL, Brandon Pratt R, Maurel C, Lachenbruch B, Zwiazek J (2012) New research on plant-water relations examines the molecular, structural, and physiological mechanisms of plant responses to their environment. New Phytologist, 196, 345-348.

Hamerlynck EP, Huxman TE, Loik ME, Smith SD (2000) Effects of extreme high temperature, drought and elevated CO2 on photosynthesis of the Mojave Desert evergreen shrub, Larrea tridentata. Plant Ecology, 148, 183-193.

Hanninen H, Tanino K (2011) Tree seasonality in a warming climate. Trends in Plant Science, 16, 412-416.

Hereford J (2009) A Quantitative Survey of Local Adaptation and Fitness Trade-Offs. American Naturalist, 173, 579-588.

Hesse PP, Humphreys GS, Selkirk PM, Adamson DA, Gore DB, Nobes DC, Price DM, Schwenninger JL, Smith B, Tulau M, Hemmings F (2003) Late Quaternary aeolian dunes on the presently humid Blue Mountains, eastern Australia. Quaternary International, 108, 13-32.

Hoover DL, Knapp AK, Smith MD (2014) Resistance and resilience of a grassland ecosystem to climate extremes. Ecology, 95, 2646-2656.

Howe GT, Aitken SN, Neale DB, Jermstad KD, Wheeler NC, Chen TH (2003) From genotype to phenotype: unraveling the complexities of cold adaptation in forest trees. Canadian Journal of Botany, 81, 1247-1266.

Hozain MI, Salvucci ME, Fokar M, Holaday AS (2010) The differential response of photosynthesis to high temperature for a boreal and temperate Populus species relates to differences in Rubisco activation and Rubisco activase properties. Tree Physiology, 30, 32-44.

Huang G, Rymer PD, Duan H, Smith RA, Tissue DT (2015) Elevated temperature is more effective than elevated [CO2] in exposing genotypic variation in Telopea speciosissima growth plasticity: implications for woody plant populations under climate change. Global Change Biology, 21, 3800-3813.

Isebrands JG, Mcdonald EP, Kruger E, Hendrey G, Percy K, Pregitzer K, Sober J, Karnosky DF (2001) Growth responses of Populus tremuloides clones to interacting elevated carbon dioxide and tropospheric ozone. Environmental Pollution, 115, 359-371.

Jeffrey SJ, Carter JO, Moodie KB, Beswick AR (2001) Using spatial interpolation to construct a comprehensive archive of Australian climate data. Environmental Modelling & Software, 16, 309-330.

Karnosky DF (2003) Impacts of elevated atmospheric CO2 on forest trees and forest ecosystems: knowledge gaps. Environment International, 29, 161-169.

Karnosky DF, Pregitzer KS, Zak DR, Kubiske ME, Hendrey GR, Weinstein D, Nosal M, Percy KE (2005) Scaling ozone responses of forest trees to the ecosystem level in a changing climate. Plant, Cell & Environment, 28, 965-981.

Kattge J, Knorr W (2007) Temperature acclimation in a biochemical model of photosynthesis: a reanalysis of data from 36 species. Plant, Cell & Environment, 30, 1176-1190.

Kawecki TJ (2008) Adaptation to Marginal Habitats. Annual Review of Ecology Evolution and Systematics, 39, 321-342.

Kharin VV, Zwiers FW, Zhang XB, Hegerl GC (2007) Changes in temperature and precipitation extremes in the IPCC ensemble of global coupled model simulations. Journal of Climate, 20, 1419-1444.

Kolb PF, Robberecht R (1996) High temperature and drought stress effects on survival of Pinus ponderosa seedlings. Tree Physiology, 16, 665-672.

Kubiske ME, Quinn VS, Heilman WE, Mcdonald EP, Marquardt PE, Teclaw RM, Friend AL, Karnosky DF (2006) Interannual climatic variation mediates elevated CO2 and O3 effects on forest growth. Global Change Biology, 12, 1054-1068.

Kubiske ME, Quinn VS, Marquardt PE, Karnosky DF (2007) Effects of elevated atmospheric CO2 and/or O3 on intra- and interspecific competitive ability of aspen. Plant Biology, 9, 342-355.

Larcher W (2003) Physiological plant ecology: ecophysiology and stress physiology of functional groups, 4th edn. Springer Science & Business Media.

Leakey ADB, Ainsworth EA, Bernacchi CJ, Rogers A, Long SP, Ort DR (2009a) Elevated CO2 effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. Journal of Experimental Botany, 60, 2859-2876.

Leakey ADB, Ainsworth EA, Bernard SM, Markelz RJC, Ort DR, Placella SA, Rogers A, Smith MD, Sudderth EA, Weston DJ, Wullschleger SD, Yuan SH (2009b) Gene expression profiling: opening the black box of plant ecosystem responses to global change. Global Change Biology, 15, 1201-1213.

Lewis JD, Lucash M, Olszyk D, Tingey DT (2001) Seasonal patterns of photosynthesis in Douglas fir seedlings during the third and fourth year of exposure to elevated CO2 and temperature. Plant, Cell & Environment, 24, 539-548.

Lewis JD, Lucash M, Olszyk DM, Tingey DT (2002) Stomatal responses of Douglas-fir seedlings to elevated carbon dioxide and temperature during the third and fourth years of exposure. Plant, Cell & Environment, 25, 1411-1421.

Lewis JD, Smith RA, Ghannoum O, Logan BA, Phillips NG, Tissue DT (2013) Industrial-age changes in atmospheric [CO2] and temperature differentially alter responses of faster- and slower-growing Eucalyptus seedlings to short-term drought. Tree Physiology, 33, 475-488.

Lloyd J, Farquhar GD (2008) Effects of rising temperatures and [CO2] on the physiology of tropical forest trees. Philosophical Transactions of the Royal Society B-Biological Sciences, 363, 1811-1817.

Matesanz S, Gianoli E, Valladares F (2010) Global change and the evolution of phenotypic plasticity in plants. Year in Evolutionary Biology, 1206, 35-55.

Mawdsley JR, O'malley R, Ojima DS (2009) A Review of Climate-Change Adaptation Strategies for Wildlife Management and Biodiversity Conservation. Conservation Biology, 23, 1080-1089.

Mckenzie D, Hessl AE, Peterson DL (2001) Recent growth of conifer species of western North America: assessing spatial patterns of radial growth trends. Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere, 31, 526-538.

Mclean EH, Prober SM, Stock WD, Steane DA, Potts BM, Vaillancourt RE, Byrne M (2014) Plasticity of functional traits varies clinally along a rainfall gradient in Eucalyptus tricarpa. Plant, Cell & Environment, 37, 1440-1451.

Meehl GA, Tebaldi C (2004) More intense, more frequent, and longer lasting heat waves in the 21st century. Science, 305, 994-997.

Melillo JM, Mcguire AD, Kicklighter DW, Moore B, Vorosmarty CJ, Schloss AL (1993) Global Climate-Change and Terrestrial Net Primary Production. Nature, 363, 234-240.

Mitchell PJ, O'Grady AP, Tissue DT, White DA, Ottenschlaeger ML, Pinkard EA (2013) Drought response strategies define the relative contributions of hydraulic dysfunction and carbohydrate depletion during tree mortality. New Phytologist, 197, 862-872.

Mittler R (2006) Abiotic stress, the field environment and stress combination. Trends in Plant Science, 11, 15-19.

Mohan JE, Clark JS, Schlesinger WH (2004) Genetic variation in germination, growth, and survivorship of red maple in response to subambient through elevated atmospheric CO2. Global Change Biology, 10, 233-247.

Moise AF, Hudson DA (2008) Probabilistic predictions of climate change for Australia and southern Africa using the reliability ensemble average of IPCCCMIP3 model simulations. Journal of Geophysical Research: Atmospheres, 113, D15113, doi: 10.11029/12007JD009250.

Monclus R, Dreyer E, Villar M, Delmotte FM, Delay D, Petit JM, Barbaroux C, Thiec D, Brechet C, Brignolas F (2006) Impact of drought on productivity and water use efficiency in 29 genotypes of Populus deltoides x Populus nigra. New Phytologist, 169, 765-777.

Moran EV, Hartig F, Bell DM (2016) Intraspecific trait variation across scales: implications for understanding global change responses. Global Change Biology, 22, 137-150.

Morison JIL (1993) Response of plants to CO2 under water limited conditions. Vegetatio, 104-105, 193-209.

Morison JIL, Lawlor DW (1999) Interactions between increasing CO2 concentration and temperature on plant growth. Plant, Cell & Environment, 22, 659-682.

Muller B, Pantin F, Génard M, Turc O, Freixes S, Piques M, Gibon Y (2011) Water deficits uncouple growth from photosynthesis, increase C content, and modify the relationships between C and growth in sink organs. Journal of Experimental Botany, 62, 1715-1729.

Nakagawa S, Schielzeth H (2013) A general and simple method for obtaining R2 from generalized linear mixed-effects models. Methods in Ecology and Evolution, 4, 133-142.

Nemani RR, Keeling CD, Hashimoto H, Jolly WM, Piper SC, Tucker CJ, Myneni RB, Running SW (2003) Climate-driven increases in global terrestrial net primary production from 1982 to 1999. Science, 300, 1560-1563.

Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U, Poot P, Purugganan MD, Richards CL, Valladares F, Van Kleunen M (2010) Plant phenotypic plasticity in a changing climate. Trends in Plant Science, 15, 684-692.

Nixon P (1997) The Waratah, 2nd edn. Kangaroo Press, Sydney.

Noormets A, Mcdonald E, Dickson R, Kruger E, Sôber A, Isebrands J, Karnosky D (2001) The effect of elevated carbon dioxide and ozone on leaf- and branch-level photosynthesis and potential plant-level carbon gain in aspen. Trees, 15, 262-270.

Norby RJ, Delucia EH, Gielen B, Calfapietra C, Giardina CP, King JS, Ledford J, Mccarthy HR, Moore DJP, Ceulemans R, De Angelis P, Finzi AC, Karnosky DF, Kubiske ME, Lukac M, Pregitzer KS, Scarascia-Mugnozza GE, Schlesinger WH, Oren R (2005) Forest response to elevated CO2 is conserved across a broad range of productivity. Proceedings of the National Academy of Sciences of the United States of America, 102, 18052-18056.

Nowak RS, Ellsworth DS, Smith SD (2004) Functional responses of plants to elevated atmospheric CO2 - do photosynthetic and productivity data from FACE experiments support early predictions? New Phytologist, 162, 253-280.

Oishi AC, Oren R, Novick K, Palmroth S, Katul G (2010) Interannual Invariability of Forest Evapotranspiration and Its Consequence to Water Flow Downstream. Ecosystems, 13, 421-436.

Pan Y, Birdsey RA, Fang J, Houghton R, Kauppi PE, Kurz WA, Phillips OL, Shvidenko A, Lewis SL, Canadell JG, Ciais P, Jackson RB, Pacala SW, Mcguire AD, Piao S, Rautiainen A, Sitch S, Hayes D (2011) A large and persistent carbon sink in the world's forests. Science, 333, 988-993.

Pearce K, Holper PN, Hopkins M, Bouma WJ, Whetton P, Hennessy KJ, Power SB (2007) Climate Change in Australia: technical report 2007, CSIRO Marine and Atmospheric Research.

Peltola H, Kilpelainen A, Kellomaki S (2002) Diameter growth of Scots pine (Pinus sylvestris) trees grown at elevated temperature and carbon dioxide concentration under boreal conditions. Tree Physiology, 22, 963-972.

Perkins SE, Alexander LV (2013) On the Measurement of Heat Waves. Journal of Climate, 26, 4500-4517.

Perry LG, Shafroth PB, Blumenthal DM, Morgan JA, Lecain DR (2013) Elevated CO2 does not offset greater water stress predicted under climate change for native and exotic riparian plants. New Phytologist, 197, 532-543.

Pinheiro J, Bates D, Debroy S, Sarkar D, R Core Team (2016) nlme: Linear and Nonlinear Mixed Effects Models, R Package version 3.1-126, http://CRAN.R-project.org/package=nlme.

Pittock AB (2003) Climate change: an Australian guide to the science and potential impacts, Australian Greenhouse Office Canberra.

Poorter H, Pérez-Soba M (2001) The growth response of plants to elevated CO2 under non-optimal environmental conditions. Oecologia, 129, 1-20.

Prudhomme C, Giuntoli I, Robinson EL, Clark DB, Arnell NW, Dankers R, Fekete BM, Franssen W, Gerten D, Gosling SN, Hagemann S, Hannah DM, Kim H, Masaki Y, Satoh Y, Stacke T, Wada Y, Wisser D (2014) Hydrological droughts in the 21st

century, hotspots and uncertainties from a global multimodel ensemble experiment. Proceedings of the National Academy of Sciences, 111, 3262-3267.

Rahmstorf S, Coumou D (2011) Increase of extreme events in a warming world. Proceedings of the National Academy of Sciences of the United States of America, 108, 17905-17909.

Ramirez-Valiente JA, Sanchez-Gomez D, Aranda I, Valladares F (2010) Phenotypic plasticity and local adaptation in leaf ecophysiological traits of 13 contrasting cork oak populations under different water availabilities. Tree Physiology, 30, 618-627.

Rehfeldt GE, Ying CC, Spittlehouse DL, Hamilton DA (1999) Genetic responses to climate in Pinus contorta: Niche breadth, climate change, and reforestation. Ecological Monographs, 69, 375-407.

Reichstein M, Bahn M, Ciais P, Frank D, Mahecha MD, Seneviratne SI, Zscheischler J, Beer C, Buchmann N, Frank DC, Papale D, Rammig A, Smith P, Thonicke K, Van Der Velde M, Vicca S, Walz A, Wattenbach M (2013) Climate extremes and the carbon cycle. Nature, 500, 287-295.

Reichstein M, Ciais P, Papale D, Valentini R, Running S, Viovy N, Cramer W, Granier A, Ogee J, Allard V, Aubinet M, Bernhofer C, Buchmann N, Carrara A, Grunwald T, Heimann M, Heinesch B, Knohl A, Kutsch W, Loustau D, Manca G, Matteucci G, Miglietta F, Ourcival JM, Pilegaard K, Pumpanen J, Rambal S, Schaphoff S, Seufert G, Soussana JF, Sanz MJ, Vesala T, Zhao M (2007) Reduction of ecosystem productivity and respiration during the European summer 2003 climate anomaly: a joint flux tower, remote sensing and modelling analysis. Global Change Biology, 13, 634-651.

Riikonen J, Kets K, Darbah J, Oksanen E, Sober A, Vapaavuori E, Kubiske ME, Nelson N, Karnosky DF (2008) Carbon gain and bud physiology in Populus tremuloides and Betula papyrifera grown under long-term exposure to elevated concentrations of CO2 and O3. Tree Physiology, 28, 243-254.

Robinson EA, Ryan GD, Newman JA (2012) A meta-analytical review of the effects of elevated CO2 on plant–arthropod interactions highlights the importance of interacting environmental and biological variables. New Phytologist, 194, 321-336.

Robson TM, Sanchez-Gomez D, Cano FJ, Aranda I (2012) Variation in functional leaf traits among beech provenances during a Spanish summer reflects the differences in their origin. Tree Genetics & Genomes, 8, 1111-1121.

Rossetto M, Thurlby K, Offord C, Allen C, Weston P (2011) The impact of distance and a shifting temperature gradient on genetic connectivity across a heterogeneous landscape. BMC Evolutionary Biology, 11, 1-11.

Sage RF, Kubien DS (2007) The temperature response of C3 and C4 photosynthesis. Plant, Cell & Environment, 30, 1086-1106.

Sage RF, Way DA, Kubien DS (2008) Rubisco, Rubisco activase, and global climate change. Journal of Experimental Botany, 59, 1581-1595.

Sala A, Woodruff DR, Meinzer FC (2012) Carbon dynamics in trees: feast or famine? Tree Physiology, 32, 764-775.

Sala OE, Chapin FS, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, Huber-Sanwald E, Huenneke LF, Jackson RB, Kinzig A, Leemans R, Lodge DM, Mooney HA, Oesterheld M, Poff NL, Sykes MT, Walker BH, Walker M, Wall DH (2000) Biodiversity - Global biodiversity scenarios for the year 2100. Science, 287, 1770-1774.

Savolainen O, Pyhajarvi T, Knurr T (2007) Gene flow and local adaptation in trees. Annual Review of Ecology Evolution and Systematics, 38, 595-619.

Saxe H, Cannell MGR, Johnsen B, Ryan MG, Vourlitis G (2001) Tree and forest functioning in response to global warming. New Phytologist, 149, 369-399.

Saxe H, Ellsworth DS, Heath J (1998) Tree and forest functioning in an enriched CO2 atmosphere. New Phytologist, 139, 395-436.

Schimel DS, House JI, Hibbard KA, Bousquet P, Ciais P, Peylin P, Braswell BH, Apps MJ, Baker D, Bondeau A, Canadell J, Churkina G, Cramer W, Denning AS, Field CB, Friedlingstein P, Goodale C, Heimann M, Houghton RA, Melillo JM, Moore B, 3rd, Murdiyarso D, Noble I, Pacala SW, Prentice IC, Raupach MR, Rayner PJ, Scholes RJ, Steffen WL, Wirth C (2001) Recent patterns and mechanisms of carbon exchange by terrestrial ecosystems. Nature, 414, 169-172.

Schlichting CD (1986) The Evolution of Phenotypic Plasticity in Plants. Annual Review of Ecology and Systematics, 17, 667-693.

Seneweera S, Norton RM (2011) Plant Responses to Increased Carbon Dioxide. In: Crop Adaptation to Climate Change. (eds Yadav SS, Redden RJ, Hatfield JL, Lotze-Campen H, Hall AE) pp 198-217. Oxford, United Kingdom, Wiley-Blackwell.

Sharkey TD, Bernacchi CJ, Farquhar GD, Singsaas EL (2007) Fitting photosynthetic carbon dioxide response curves for C3 leaves. Plant, Cell & Environment, 30, 1035-1040.

Silva FCE, Shvaleva A, Maroco JP, Almeida MH, Chaves MM, Pereira JS (2004) Responses to water stress in two Eucalyptus globulus clones differing in drought tolerance. Tree Physiology, 24, 1165-1172.

Silva JCE, Potts BM, Dutkowski GW (2006) Genotype by environment interaction for growth of Eucalyptus globulus in Australia. Tree Genetics & Genomes, 2, 61-75.

Smith AM, Stitt M (2007) Coordination of carbon supply and plant growth. Plant, Cell & Environment, 30, 1126-1149.

Smith MD (2011) An ecological perspective on extreme climatic events: a synthetic definition and framework to guide future research. Journal of Ecology, 99, 656-663.

Solomon S, Plattner GK, Knutti R, Friedlingstein P (2009) Irreversible climate change due to carbon dioxide emissions. Proceedings of the National Academy of Sciences of the United States of America, 106, 1704-1709.

Stefanon M, Drobinski P, D'andrea F, Lebeaupin-Brossier C, Bastin S (2014) Soil moisture-temperature feedbacks at meso-scale during summer heat waves over Western Europe. Climate Dynamics, 42, 1309-1324.

Stitt M, Krapp A (1999) The interaction between elevated carbon dioxide and nitrogen nutrition: the physiological and molecular background. Plant, Cell & Environment, 22, 583-621.

Sultan SE (2000) Phenotypic plasticity for plant development, function and life history. Trends in Plant Science, 5, 537-542.

Taylor G, Tallis MJ, Giardina CP, Percy KE, Miglietta F, Gupta PS, Gioli B, Calfapietra C, Gielen B, Kubiske ME, Scarascia-Mugnozza GE, Kets K, Long SP, Karnosky DF (2008) Future atmospheric CO2 leads to delayed autumnal senescence. Global Change Biology, 14, 264-275.

Tebaldi C, Hayhoe K, Arblaster JM, Meehl GA (2006) Going to the extremes. Climatic Change, 79, 185-211.

Teskey R, Wertin T, Bauweraerts I, Ameye M, Mcguire MA, Steppe K (2015) Responses of tree species to heat waves and extreme heat events. Plant, Cell & Environment, 38, 1699-1712.

Teskey RO, Will RE (1999) Acclimation of loblolly pine (Pinus taeda) seedlings to high temperatures. Tree Physiology, 19, 519-525.

Thompson JD (1991) Phenotypic Plasticity as a Component of Evolutionary Change. Trends in Ecology & Evolution, 6, 246-249.

Thomson AM, Riddell CL, Parker WH (2009) Boreal forest provenance tests used to predict optimal growth and response to climate change: 2. Black spruce. Canadian Journal of Forest Research-Revue Canadianne De Recherche Forestiere, 39, 143-153.

Tjoelker MG, Oleksyn J, Lorenc-Plucinska G, Reich PB (2009) Acclimation of respiratory temperature responses in northern and southern populations of Pinus banksiana. New Phytologist, 181, 218-229.

Trenberth KE (2011) Changes in precipitation with climate change. Climate Research, 47, 123-138.

Valladares F, Gianoli E, Gomez JM (2007) Ecological limits to plant phenotypic plasticity. New Phytologist, 176, 749-763.

Van Kleunen M, Fischer M (2005) Constraints on the evolution of adaptive phenotypic plasticity in plants. New Phytologist, 166, 49-60.

Vautard R, Yiou P, D'andrea F, De Noblet N, Viovy N, Cassou C, Polcher J, Ciais P, Kageyama M, Fan Y (2007) Summertime European heat and drought waves induced by wintertime Mediterranean rainfall deficit. Geophysical Research Letters, 34, L07711, doi: 10.01029/02006GL028001.

Wahid A, Gelani S, Ashraf M, Foolad MR (2007) Heat tolerance in plants: An overview. Environmental and Experimental Botany, 61, 199-223.

Wang D, Heckathorn SA, Wang XZ, Philpott SM (2012) A meta-analysis of plant physiological and growth responses to temperature and elevated CO2. Oecologia, 169, 1-13.

Wang TL, O'neill GA, Aitken SN (2010) Integrating environmental and genetic effects to predict responses of tree populations to climate. Ecological Applications, 20, 153-163.

Wang WX, Vinocur B, Altman A (2003) Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. Planta, 218, 1-14.

Way DA (2013) Will rising CO2 and temperatures exacerbate the vulnerability of trees to drought? Tree Physiology, 33, 775-778.

Way DA, Oren R (2010) Differential responses to changes in growth temperature between trees from different functional groups and biomes: a review and synthesis of data. Tree Physiology, 30, 669-688.

Way DA, Sage RF (2008) Thermal acclimation of photosynthesis in black spruce [Picea mariana (Mill.) BSP]. Plant, Cell & Environment, 31, 1250-1262.

Weinig C (2000) Plasticity versus canalization: Population differences in the timing of shade-avoidance responses. Evolution, 54, 441-451.

Wertin TM, Mcguire MA, Teskey RO (2010) The influence of elevated temperature, elevated atmospheric CO2 concentration and water stress on net photosynthesis of loblolly pine (Pinus taeda L.) at northern, central and southern sites in its native range. Global Change Biology, 16, 2089-2103.

Wertin TM, Mcguire MA, Teskey RO (2011) Higher growth temperatures decreased net carbon assimilation and biomass accumulation of northern red oak seedlings near the southern limit of the species range. Tree Physiology, 31, 1277-1288.

Wertin TM, Mcguire MA, Teskey RO (2012) Effects of predicted future and current atmospheric temperature and [CO2] and high and low soil moisture on gas exchange and growth of Pinus taeda seedlings at cool and warm sites in the species range. Tree Physiology, 32, 847-858.

Weston DJ, Bauerle WL (2007) Inhibition and acclimation of C3 photosynthesis to moderate heat: a perspective from thermally contrasting genotypes of Acer rabrum (red maple). Tree Physiology, 27, 1083-1092.

Weston DJ, Bauerle WL, Swire-Clark GA, Moore BD, Baird WMV (2007) Characterization of Rubisco activase from thermally contrasting genotypes of Acer rubrum (Aceraceae). American Journal of Botany, 94, 926-934.

Weston P, Crisp M (1994) Cladistic biogeography of waratahs (Proteaceae, Embothrieae) and their allies across the pacific. Australian Systematic Botany, 7, 225-249.

Will RE, Wilson SM, Zou CB, Hennessey TC (2013) Increased vapor pressure deficit due to higher temperature leads to greater transpiration and faster mortality during drought for tree seedlings common to the forest–grassland ecotone. New Phytologist, 200, 366-374.

Williams SE, Shoo LP, Isaac JL, Hoffmann AA, Langham G (2008) Towards an Integrated Framework for Assessing the Vulnerability of Species to Climate Change. PLoS Biology, 6, 2621-2626.

Wullschleger SD, Tschaplinski TJ, Norby RJ (2002) Plant water relations at elevated CO2 - implications for water-limited environments. Plant, Cell & Environment, 25, 319-331.

Yao Y, Luo Y, Huang JB, Zhao ZC (2013) Comparison of Monthly Temperature Extremes Simulated by CMIP3 and CMIP5 Models. Journal of Climate, 26, 7692-7707.

Zeppel MJB, Lewis JD, Chaszar B, Smith RA, Medlyn BE, Huxman TE, Tissue DT (2012) Nocturnal stomatal conductance responses to rising [CO2], temperature and drought. New Phytologist, 193, 929-938.

Zhao J, Hartmann H, Trumbore S, Ziegler W, Zhang Y (2013) High temperature causes negative whole-plant carbon balance under mild drought. New Phytologist, 200, 330-339.

Zhao MS, Running SW (2010) Drought-Induced Reduction in Global Terrestrial Net Primary Production from 2000 Through 2009. Science, 329, 940-943.

Zinta G, Abdelgawad H, Domagalska MA, Vergauwen L, Knapen D, Nijs I, Janssens IA, Beemster GTS, Asard H (2014) Physiological, biochemical, and genome-wide transcriptional analysis reveals that elevated CO2 mitigates the impact of combined heat wave and drought stress in Arabidopsis thaliana at multiple organizational levels. Global Change Biology, 20, 3670-3685.

Zweifel R, Zimmermann L, Zeugin F, Newbery DM (2006) Intra-annual radial growth and water relations of trees: implications towards a growth mechanism. Journal of Experimental Botany, 57, 1445-1459.